

ADNI PET Core Update

**WW ADNI
Boston
July 2013**



Current ADNI PET Data

Florbetapir

1011 Baseline scans (910 Processed)

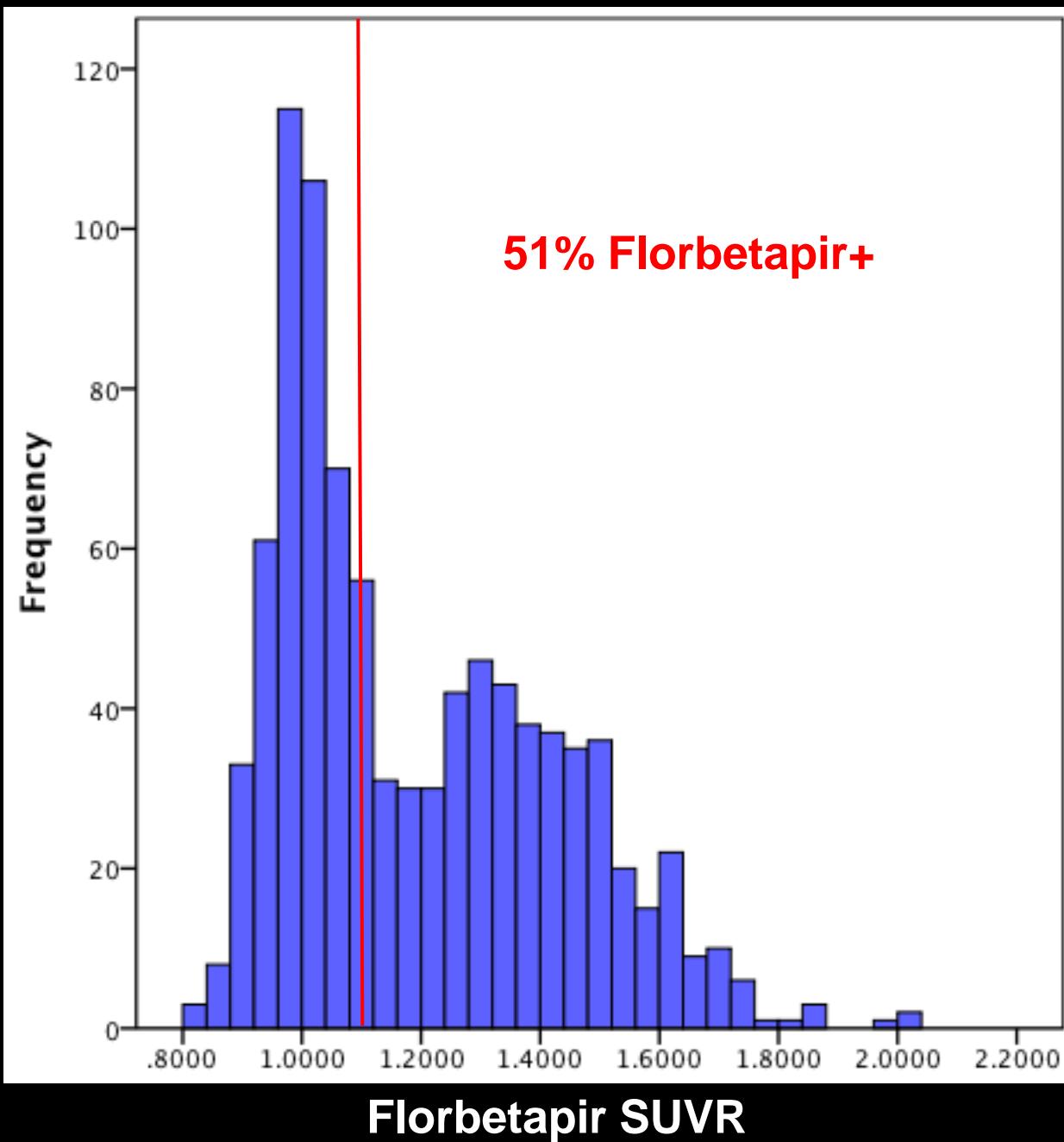
174 Follow-up scans (all processed)

39 “early frame” datasets

FDG (N = 2920)

	AD	EMCI	LMCI	N	SMC
Baseline	225	308	396	318	29
6 month	88		186	94	
12 month	76	64	202	130	
18 month			153		
24 month	62	4	149	100	
36 month			111	72	
48 month			58	57	
60 month			14	24	

Florbetapir Frequency Distribution



910 Subjects

263 Normals

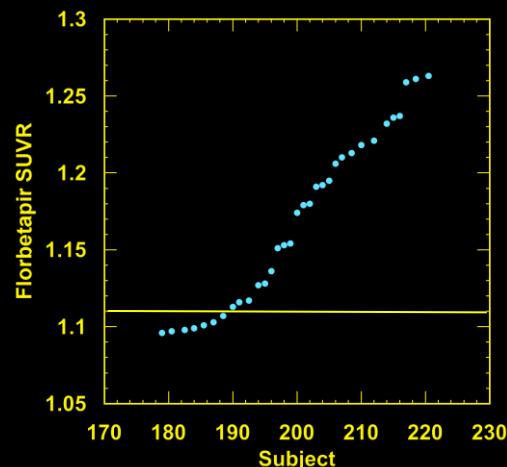
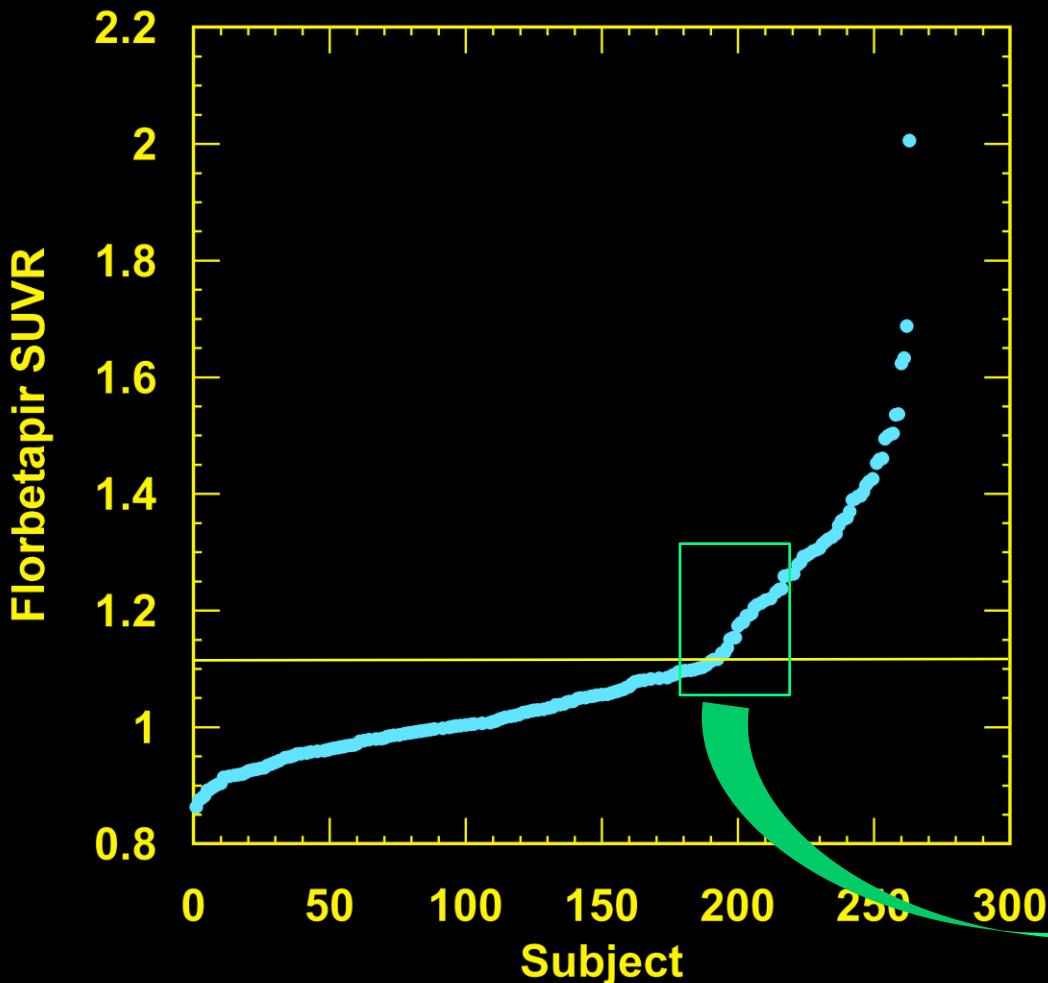
289 EMCI

212 LMCI

146 AD

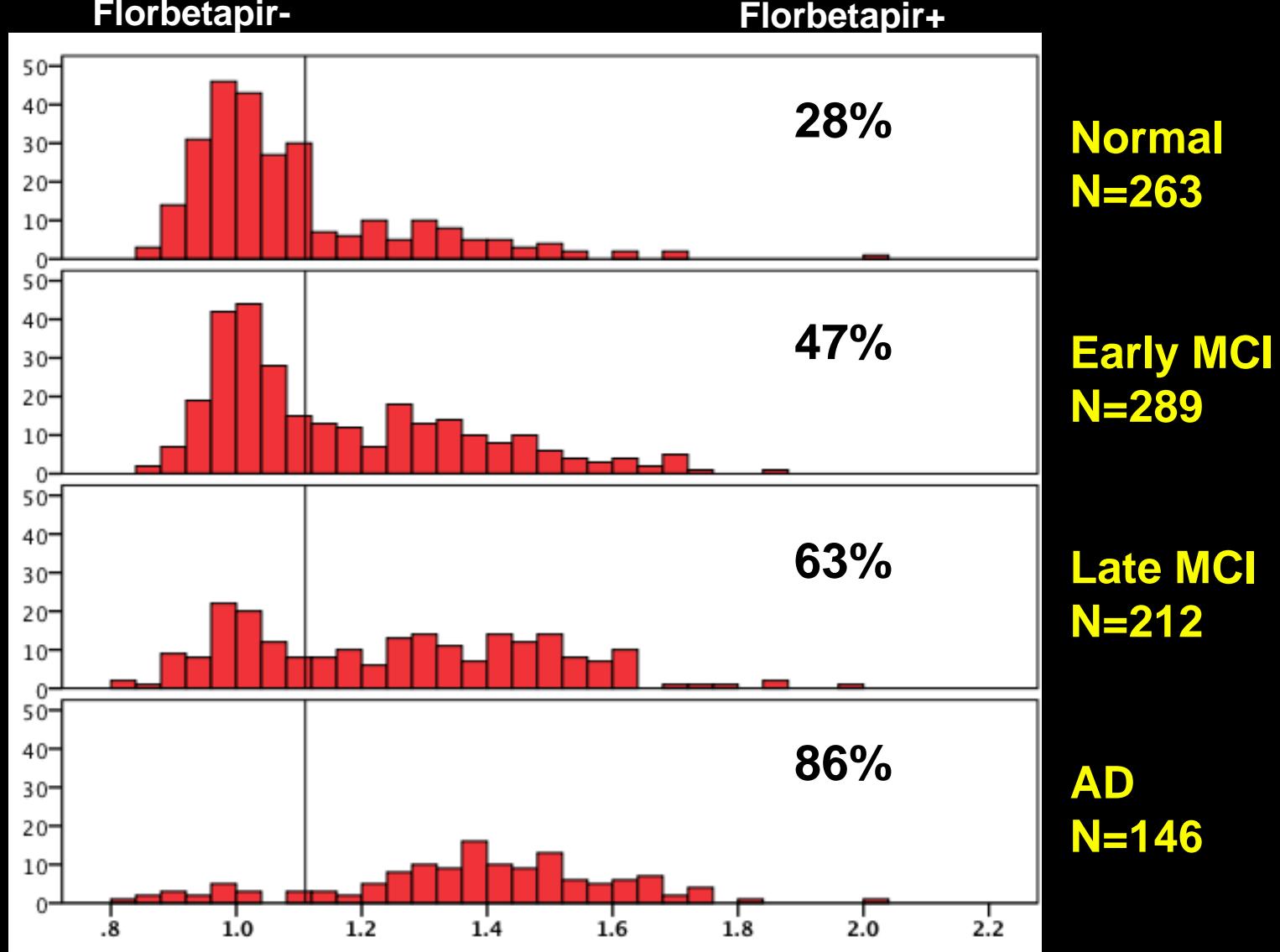
Florbetapir Threshold in Normals

1.11



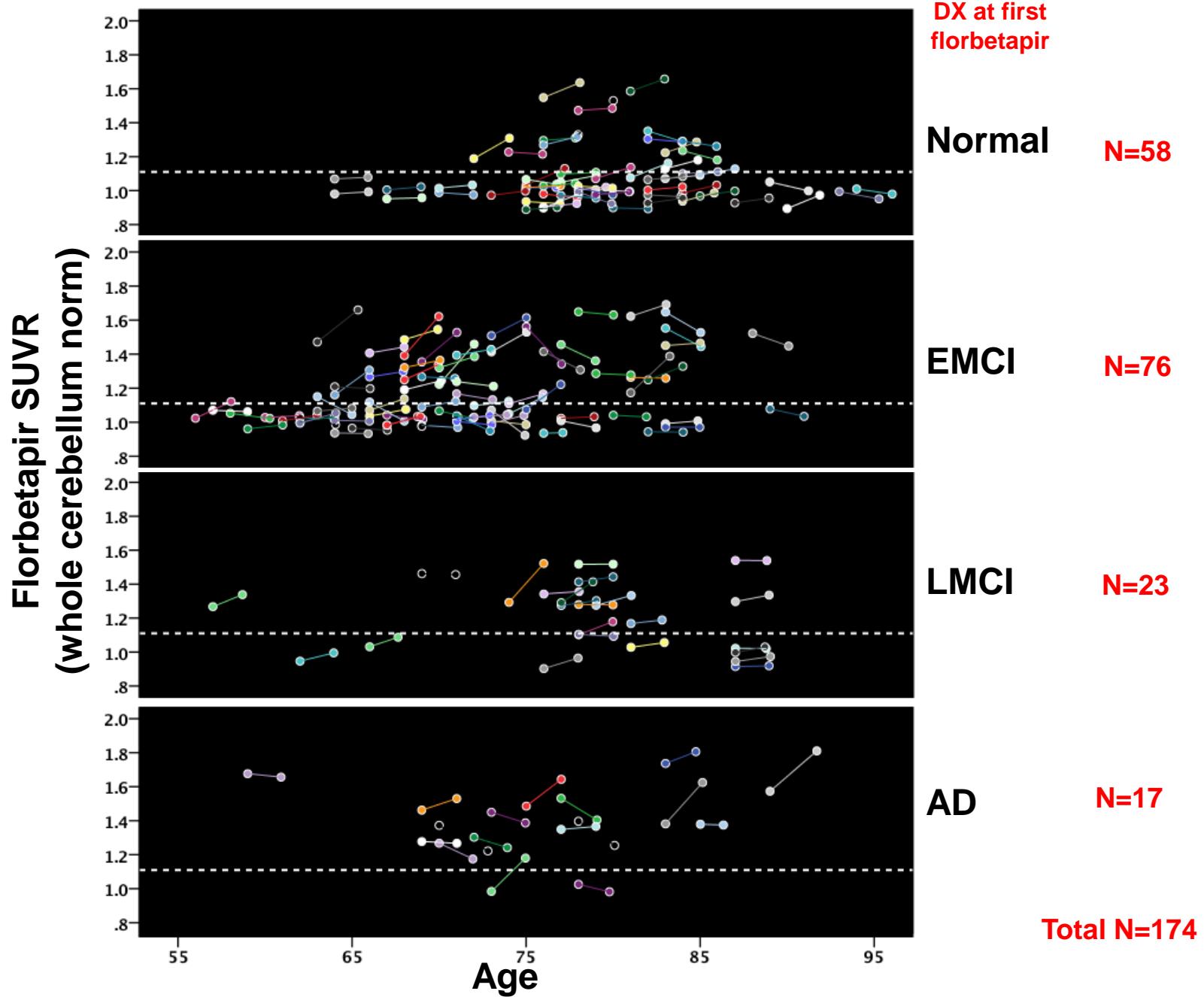
ADNI Florbetapir summary March 2013

N=910

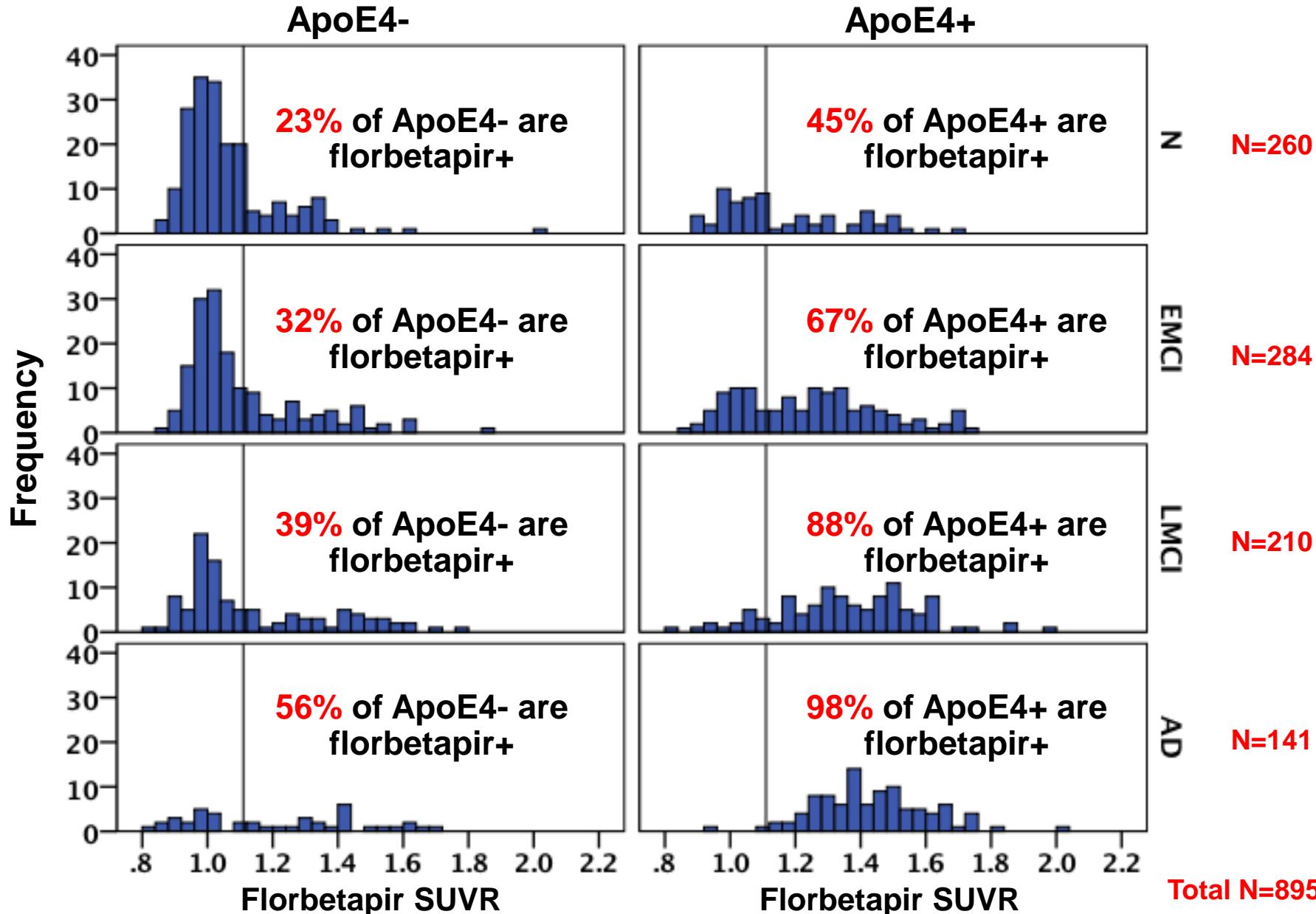


Florbetapir SUVR (Whole Cerebellum Normalization)

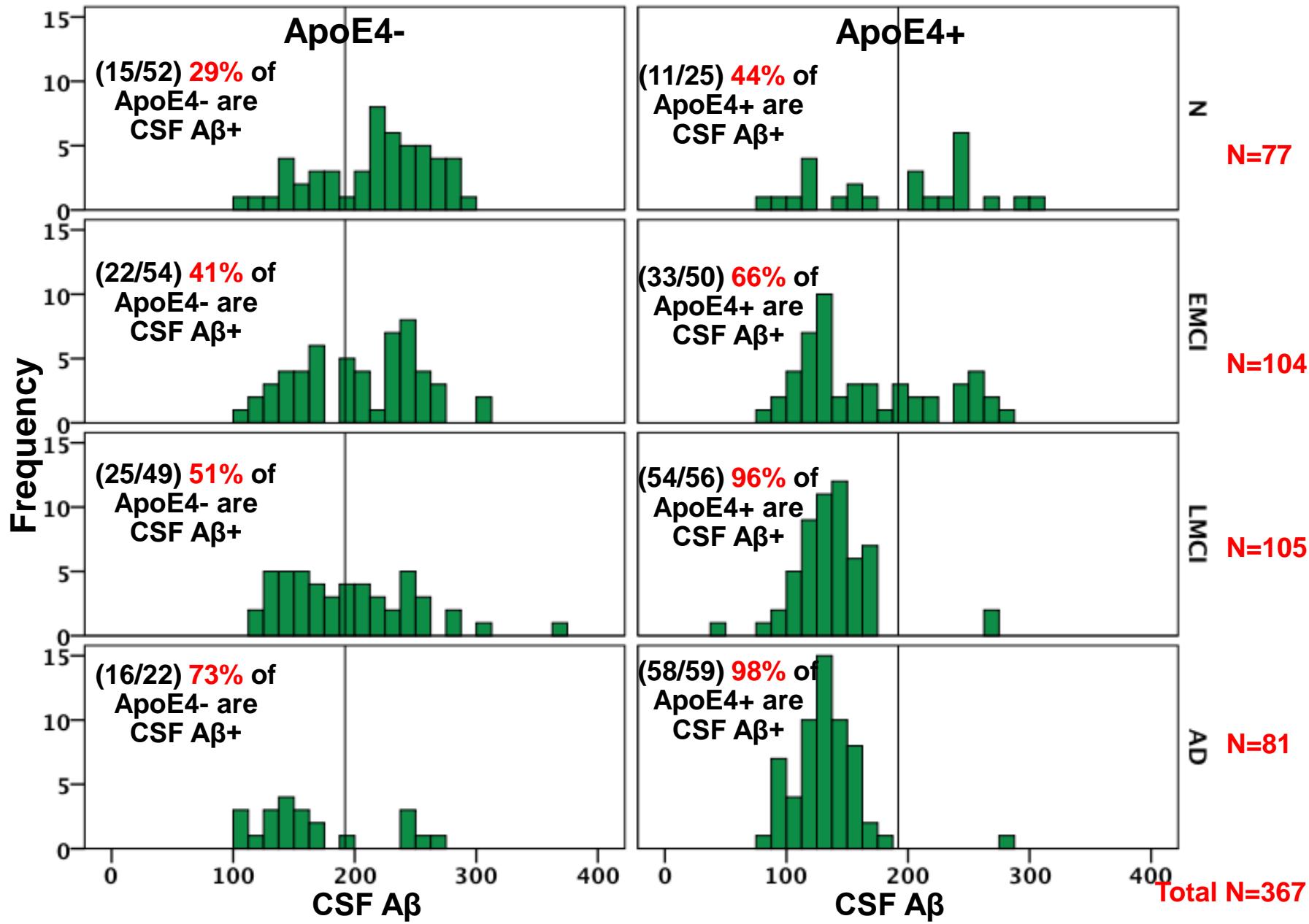
Florbetapir 2 yr change



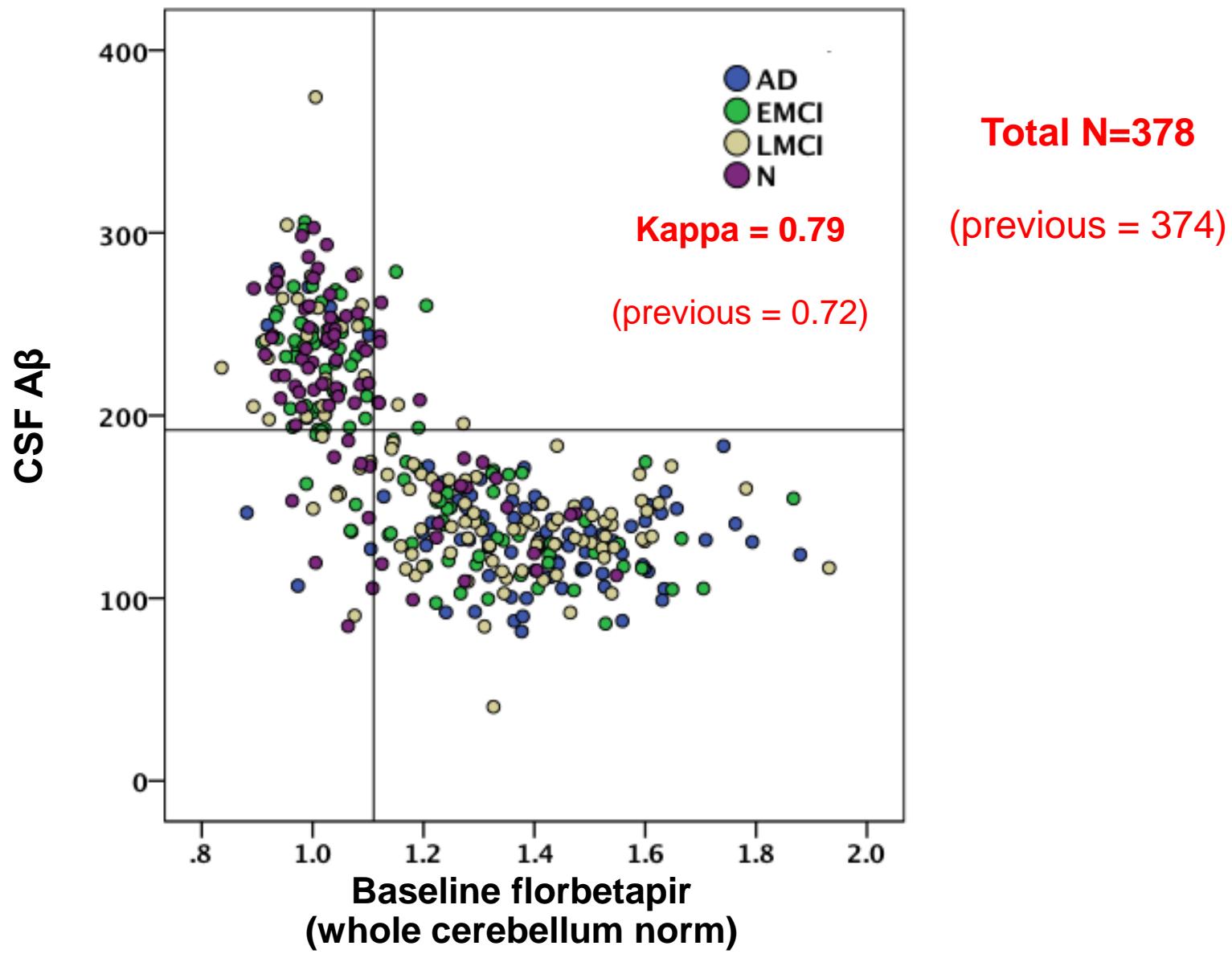
ADNI Florbetapir distribution stratified by ApoE4 status



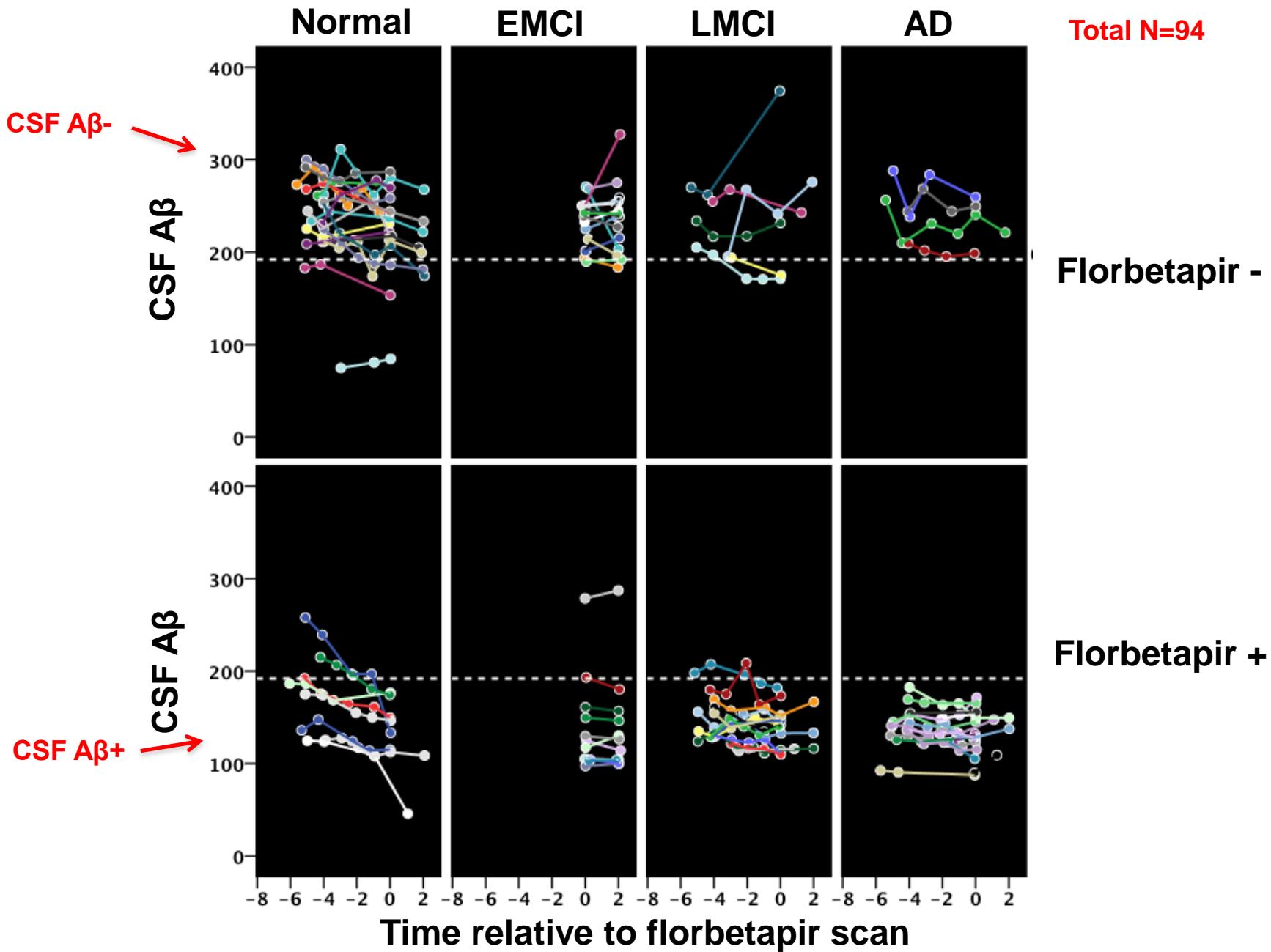
ADNI CSF A β distribution stratified by ApoE4 status



Florbetapir and CSF A β agreement

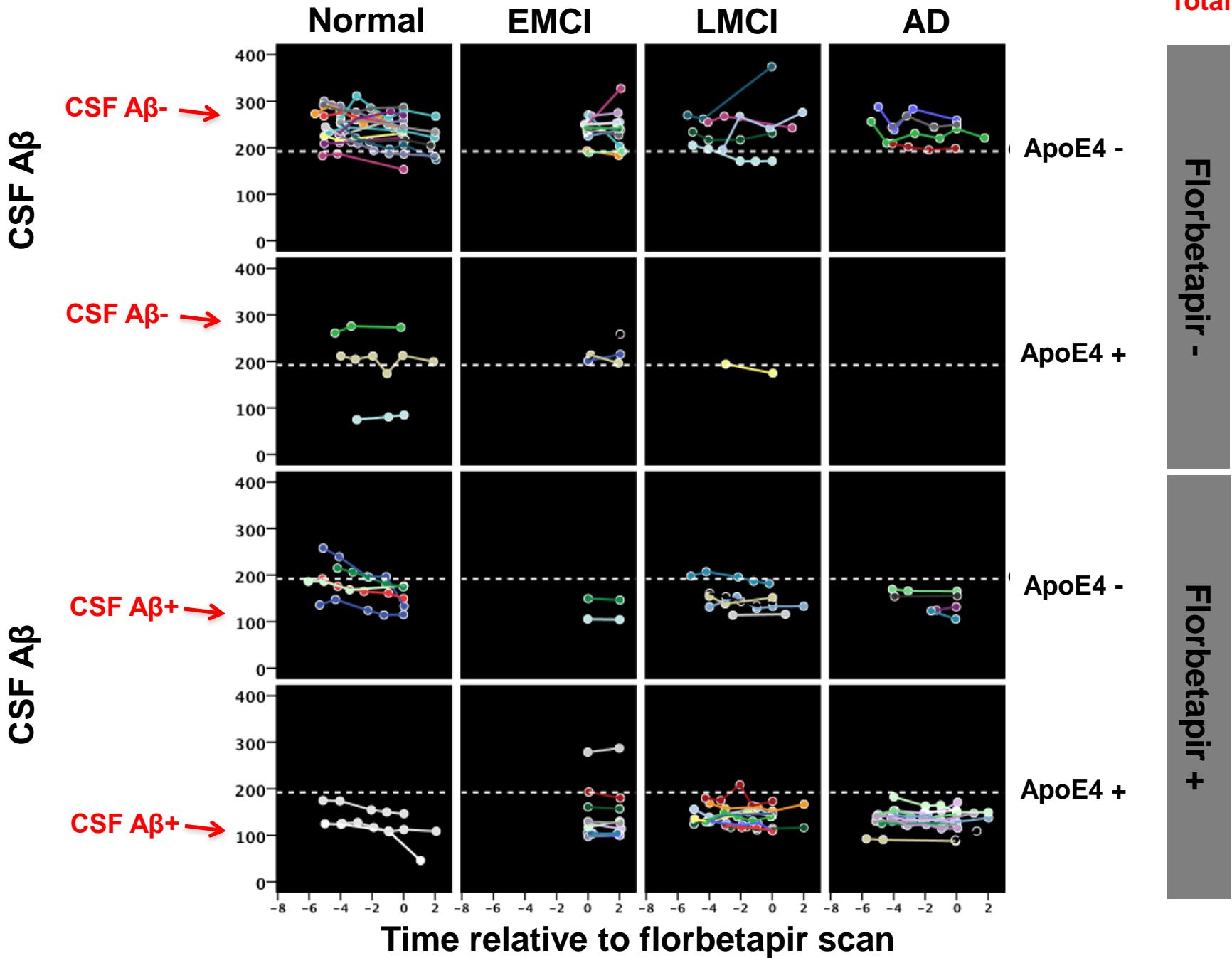


CSF change by florbetapir status



CSF change by florbetapir and APOE4 status

Total N=93



New Initiative: “Early Frames” Add on

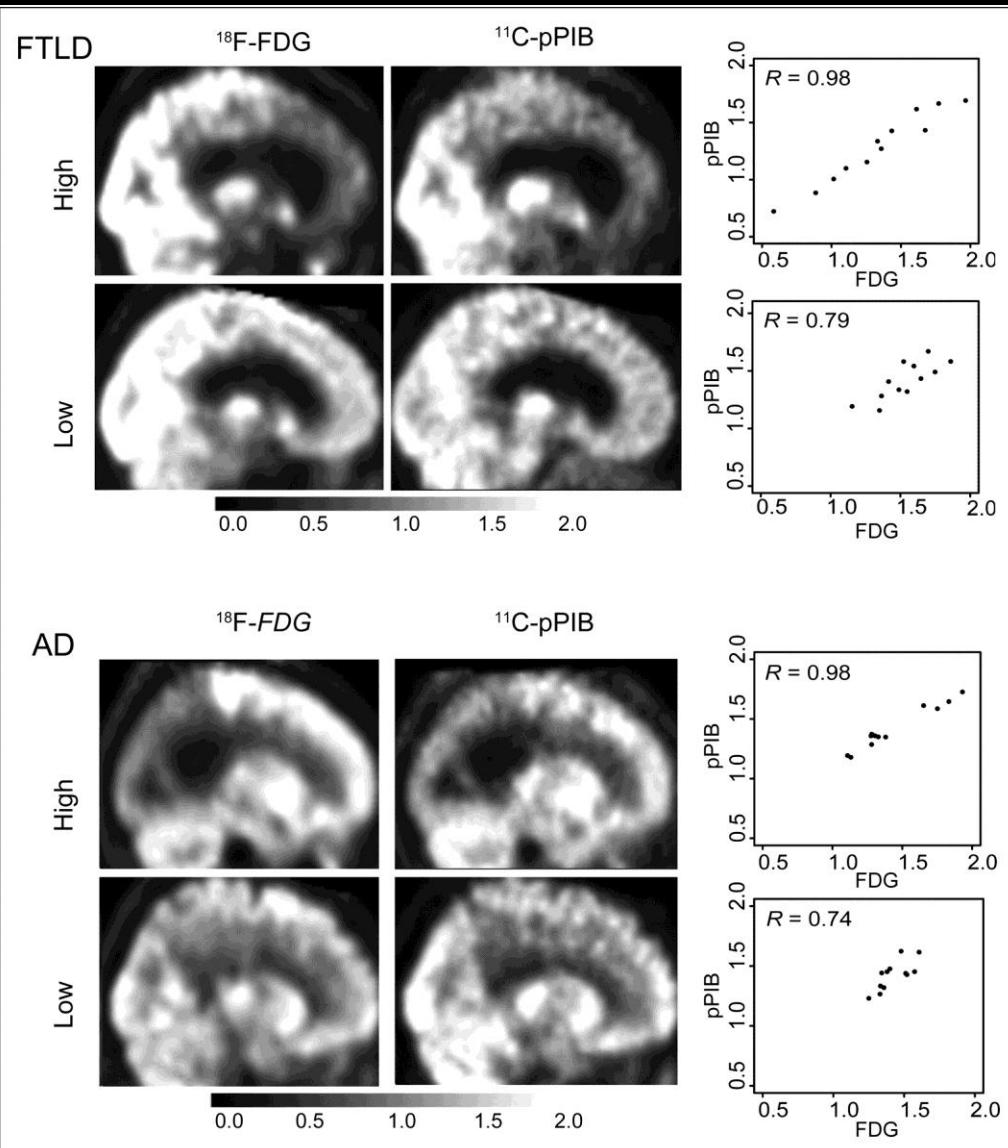
Initial uptake and clearance of highly permeable tracers reflect perfusion

Florbetapir data from injection to 50 min are not captured

Would Florbetapir data from 0-50 min provide functional information?

Compare early frame data to FDG-PET in a wide range of dementia severity/stages

Early PiB Frames vs FDG-PET



Minutes 1-8 after [¹¹C]PiB injection summed

Correlations between FDG and PiB data computed for 12 ROIs

Mean Pearson R = 0.91

(72 cases of AD or FTLD)

Study Design

Approximately 20 sites

must be capable of dynamic scanning and simultaneous injection/scan start

100 subjects: Normal, EMCI, LMCI, AD

Data collection 0-20 min, then back in scanner for the standard 50-70 min

All data treated identically to all other ADNI data

The “Centiloid’ Project

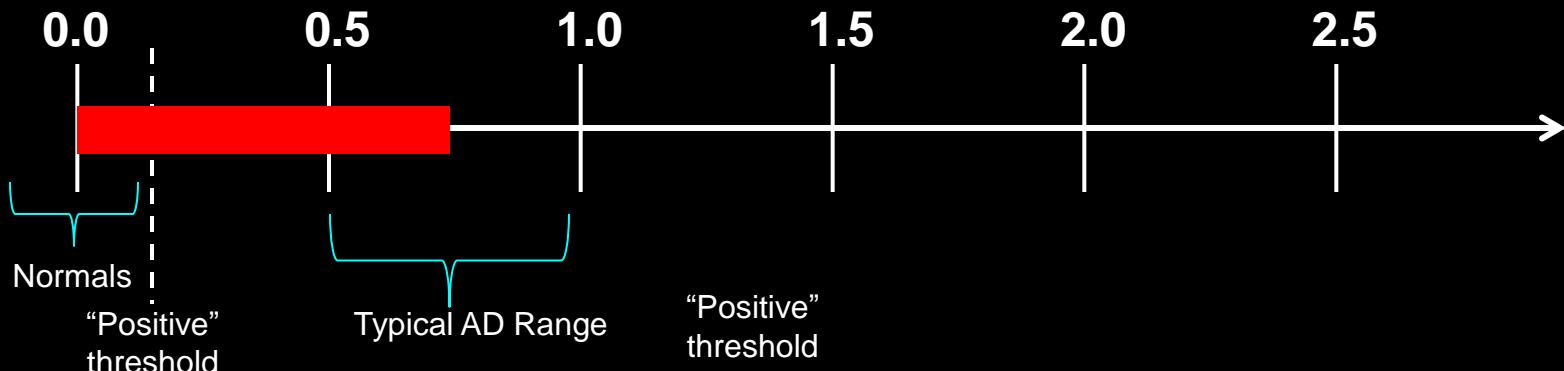
**An effort to standardize the numerical reporting
of PET amyloid tracer retention**

**Goal: A numerical value reflects the same thing
(ie, A β -, slightly A β +, very A β +, borderline)
regardless of tracer**

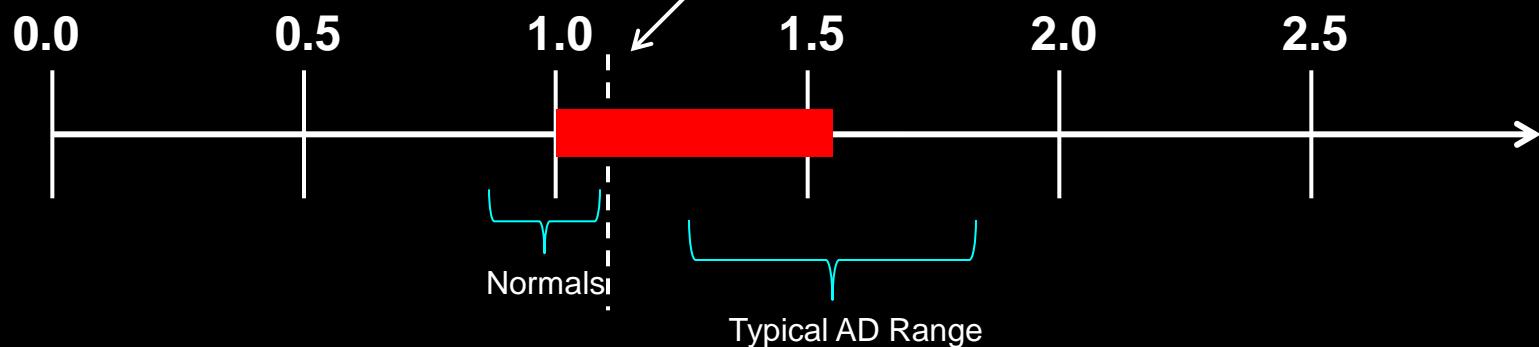
**Participants: Bill Klunk, Mike Devous, Bill Jagust,
Keith Johnson, Bob Koepp, Chet Mathis, Mark
Mintun, Mike Pontecorvo, Julie Price, Chris
Rowe, Dan Skovronsky**

Examples of Different Scales

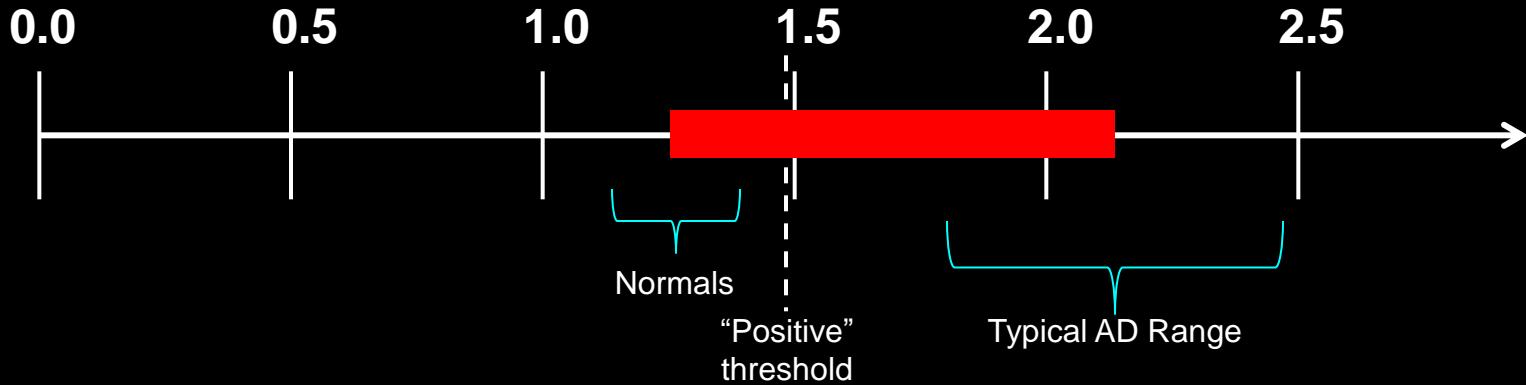
**BP
PIB**



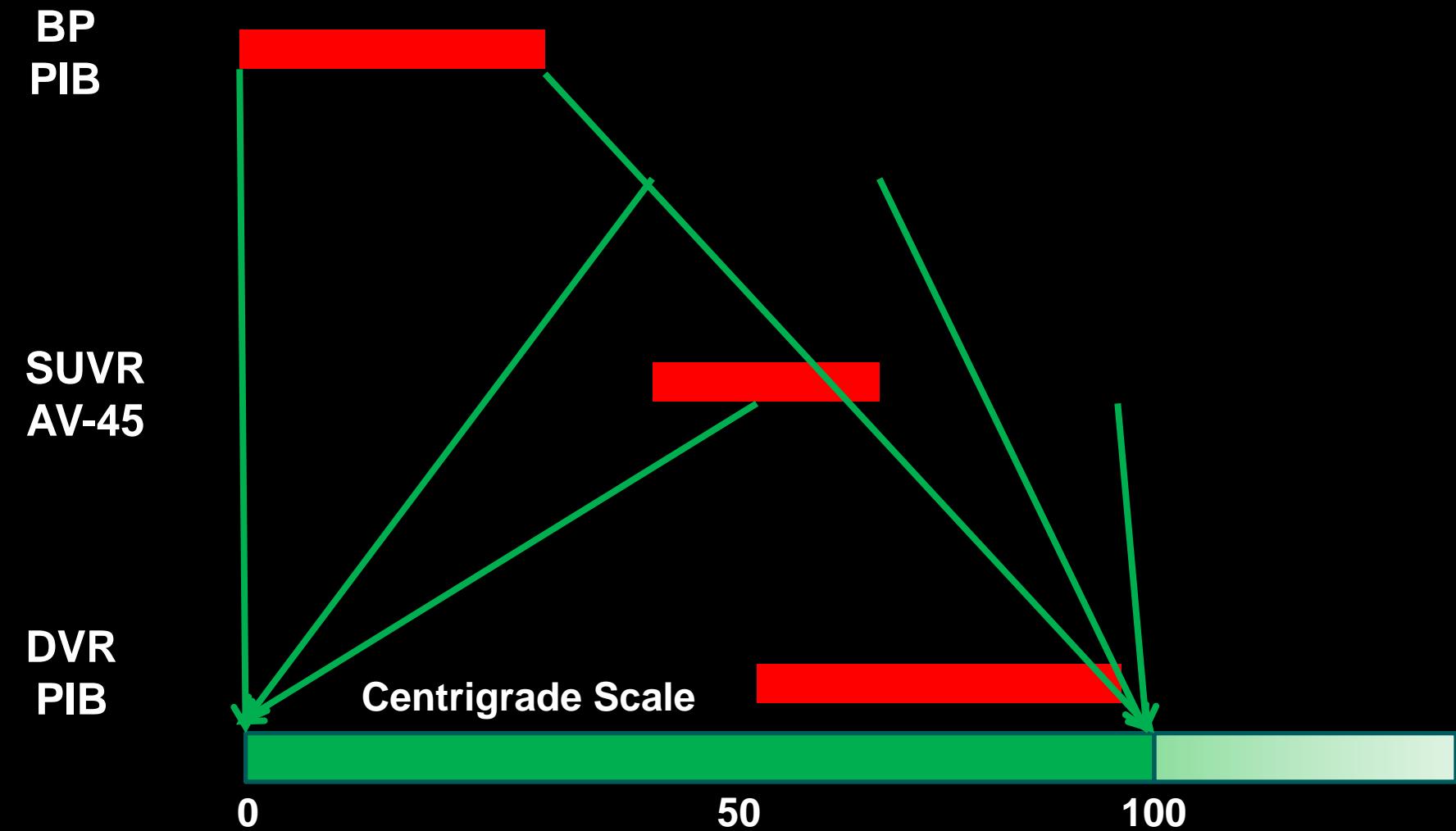
**SUVR
AV-45**



**DVR
PIB**



Conversion of Scales



The Centiloid Scale is meant to:

- Help standardize reporting across labs and tracers
- Clearly define thresholds for amyloid positivity
- Define range from “borderline” to “AD-like”
- Consistent representation of longitudinal change

The Centiloid Scale is not meant to:

- Solve all problems related to standardization
- Represent an “industry standard” requirement
- Constrain laboratories in how data are analyzed or reported

Ultimately, the amyloid imaging investigators will decide if this is a useful approach

A Brief Explanation of How it Works

Step 1: Standardization

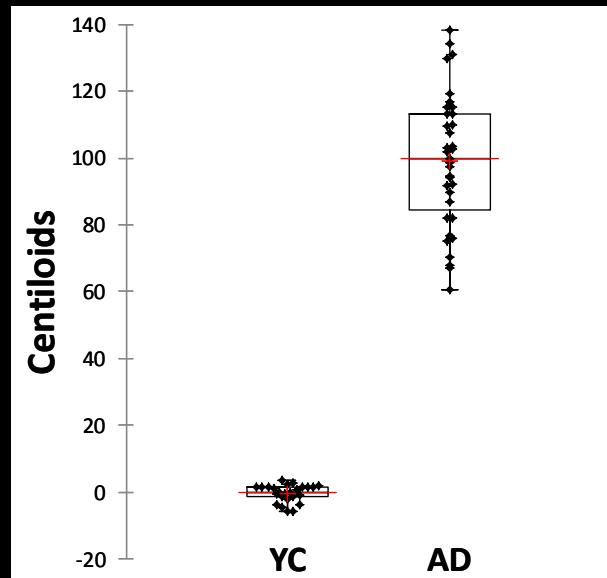
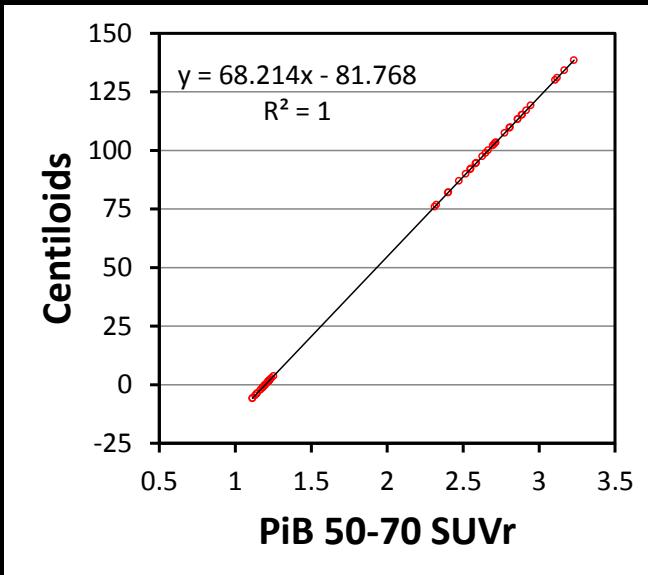
[¹¹C]PIB 50-70 min SUVR as the standard

0 anchor: Young Subjects (median)

100 anchor: Probable AD patients (median)

Standard data analysis method

Conversion of SUVR to Centiloid ($y = mx + b$)



Step 2: Calibrate new Tracer (or method)

**Collect PIB 50-70 SUVR data with new tracer data
in same subjects (including young and AD)**

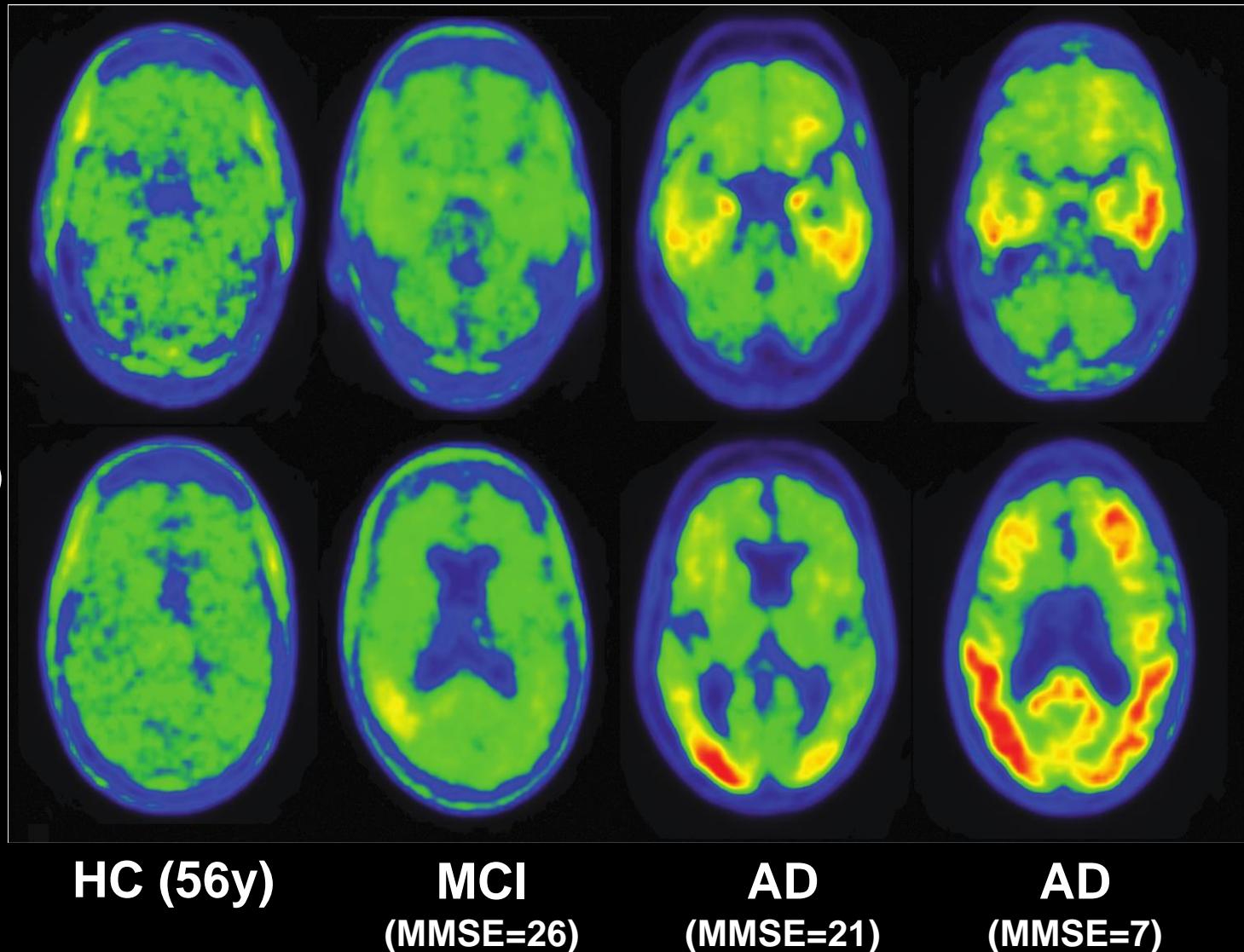
**Verification that standard analysis method works
by downloading and analyzing the standard
dataset**

Convert newly acquired PIB data to Centiloids

Convert new tracer data to Centiloids

Next Up: Tau Imaging

[¹⁸F]-T807 PET
(80-100 min p.i.)



Acknowledgements

**Susan Landau, Suzanne Baker, Bob Koeppe, Eric
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Core Leaders

Site PIs

Participants

