

Evaluating the Translational Validity of Mouse Models of Late-Onset AD (LOAD) through Deep-Phenotyping

Grant MacGregor PhD (Head)

UCI MODEL-AD Disease Modeling Project (DMP)

Gareth Howell PhD (Head)

IU/JAX MODEL-AD Disease Modeling Project (DMP)

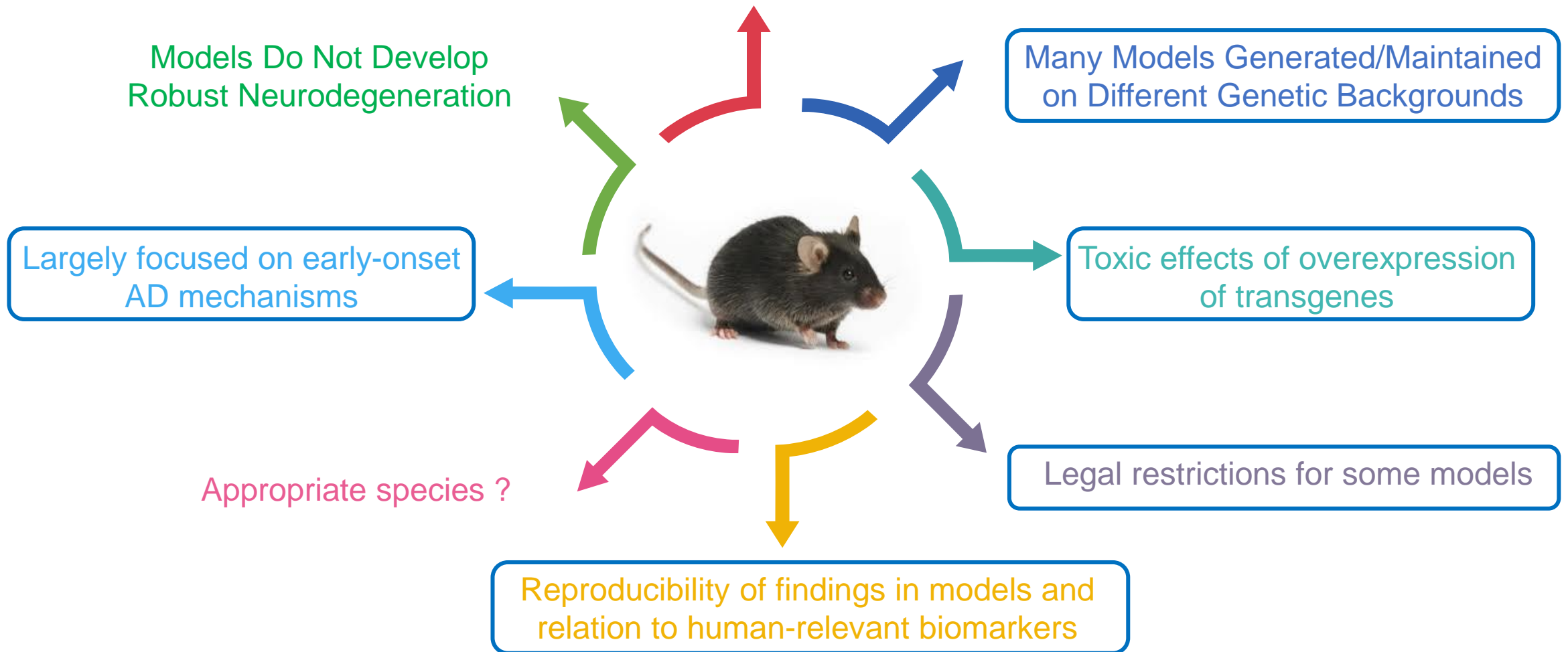
Evaluating the Translational Validity of Mouse Models of Late-Onset AD (LOAD) through Deep-Phenotyping

MODEL-AD Consortium - Disease Modeling Project (DMP)

- General strategy for model development and phenotyping platforms.
- Examples of hA β -KI (completed) and hTau-KI (in progress).
- Phenotyping of *APOE4*, *Trem2*^{R47H} models.
- Effect of mouse genetic background on development of pathology.

Concerns with Existing Animal Models of AD

Difficulties in Relating Behavioral Deficits
Observed in Mouse Models to Human AD



Using genome engineering to generate mouse models of Late-Onset Alzheimer's Disease (LOAD)

- Use CRISPR/Cas9 to introduce coding and conserved non-coding LOAD GWAS risk-variants into cognate loci in mouse genome – e.g. *Trem2*^{R47H}
- Overcomes limitations associated with -
 - Random integration of transgenes.
 - Supra-physiologic expression.
 - Lack of availability of matched negative controls.
- Accelerated production compared with previous HR / ES-cell based strategies.
- Improve reproducibility and reduce experimental variability by using consistent genetic background (C57BL/6J, initially).

Using advanced genome engineering to generate mouse models of Late-Onset Alzheimer's Disease (LOAD) – UCI DMP

- Use CRISPR/Cas9 with long (~ 2kb) ssDNA homology dependent repair (HDR) templates to introduce non-conserved LOAD GWAS risk-variants into cognate loci in mouse genome – e.g. humanizing non-conserved regions of mouse clusterin locus (*Clu*).
- Use of Recombinase Mediated Cassette Exchange (RMCE) to humanize entire loci – e.g. hTau-KI, hClu-KI.
- Generate LOAD mouse models on consistent genetic background (C57BL/6J, initially).
- Maximize researcher access to all models – available to both academics and pharma from Jackson Lab AD Mouse Model Resource, with minimal restrictions.

A humanized platform for introduction of GWAS AD-risk variants to generate mouse models of LOAD



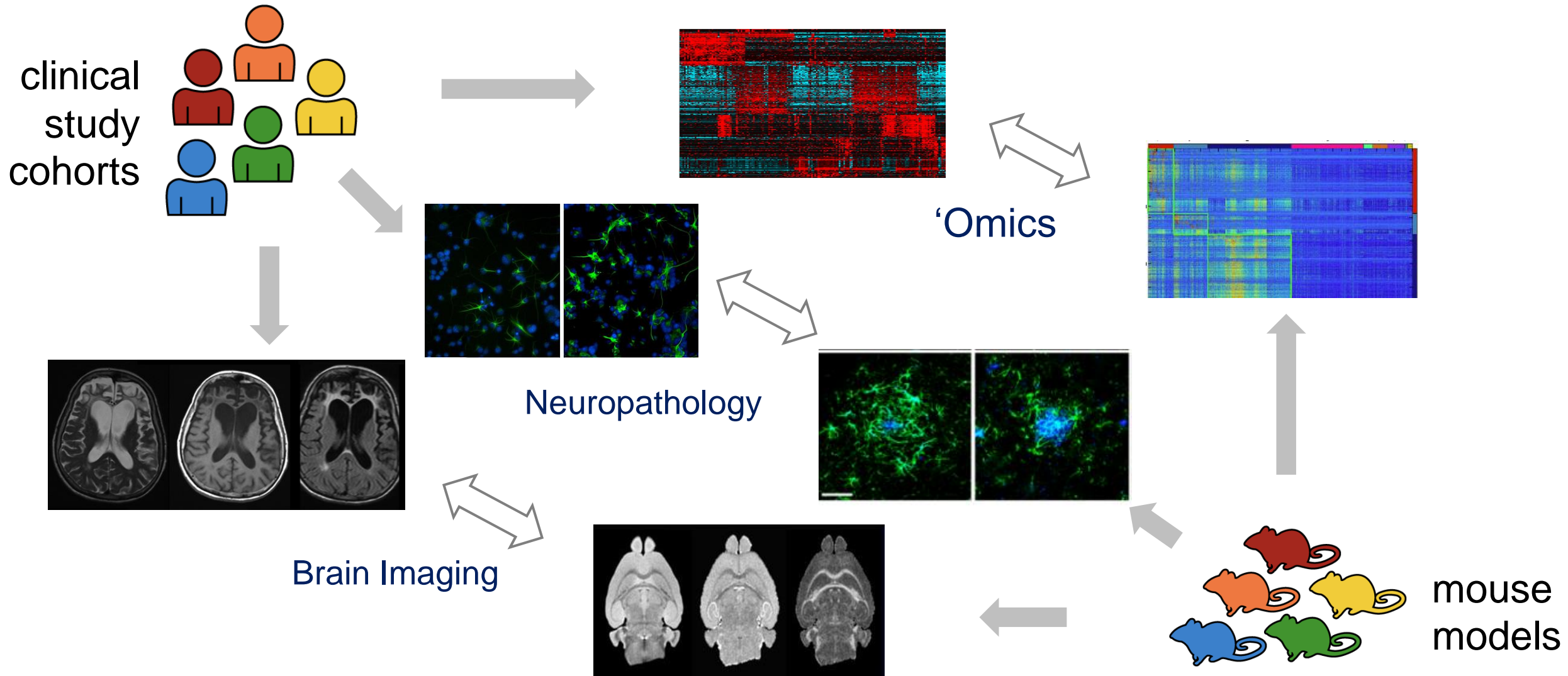
B6J. hA β -KI; APOE ϵ ⁴/ ϵ ⁴; hTau-KI
base platform

Available now In development

Long-term goal

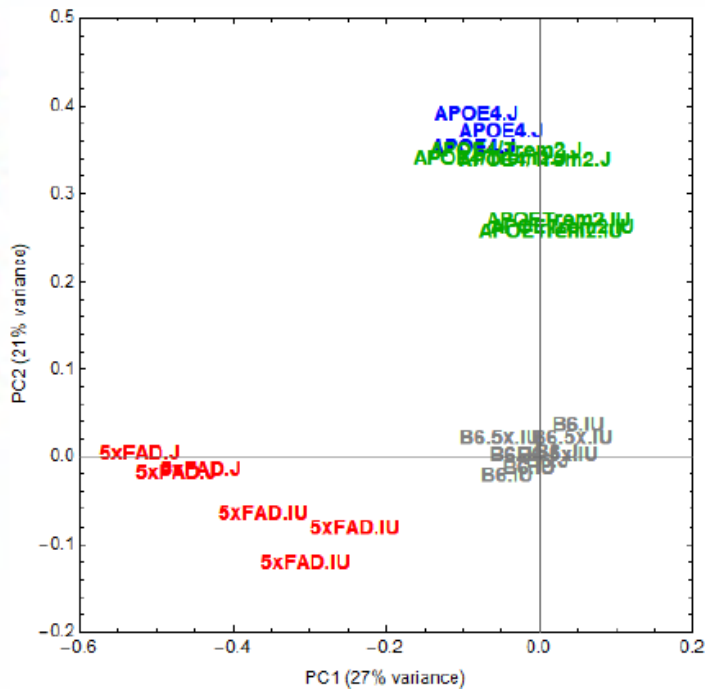
- Introduce different combinations of GWAS human LOAD risk alleles into *hA β -KI; APOE ϵ ⁴; hTau-KI* via CRISPR/Cas9 or assisted reproduction.
- Perform initial screen, then deep-phenotyping on subset to analyze effects.
 - *Trem2*^{R47H}
 - *Abca7*^{A1527G}
 - *Plcg2*^{M28L}
 - *Mthfr*^{A222V}

Goal - alignment of mouse models with clinical measures



Assessing reproducibility of findings at different sites

- Phenotype = Genes + Environment
- Harmonize environment and methodology to extent possible at each site.
- Assess reproducibility of data generated from deep-phenotyping.



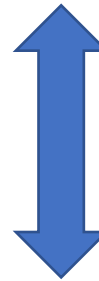
INDIANA UNIVERSITY

- 5xfAD
- APOE4
- APOE4;Trem2^{R47H}
- B6J



The Jackson Laboratory

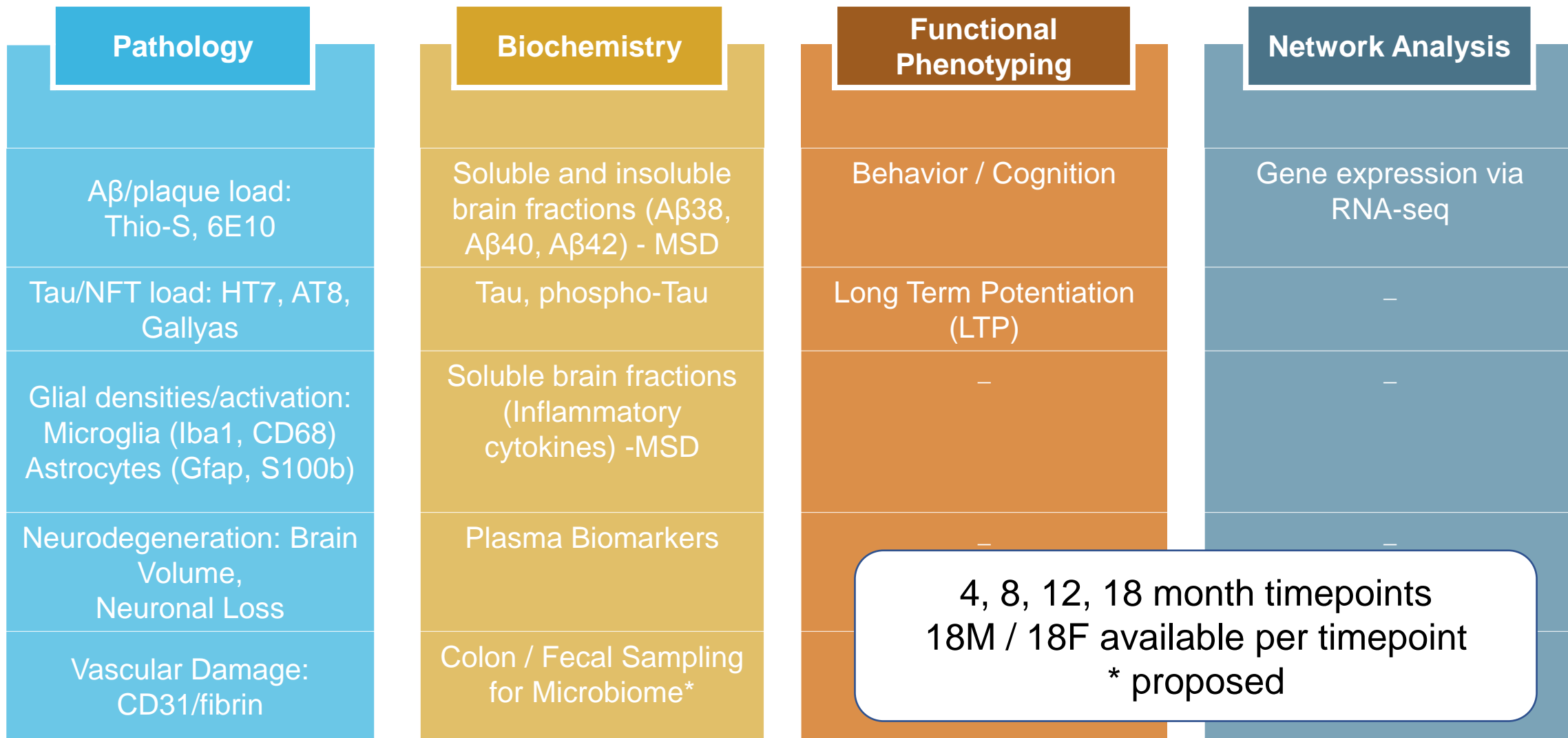
- 5xfAD
- hAβ-KI
- hAβ-KI; APOE4
- hAβ-KI; APOE4;Trem2^{R47H}
- B6J



e.g. IU v JAX mice nanoString Analysis

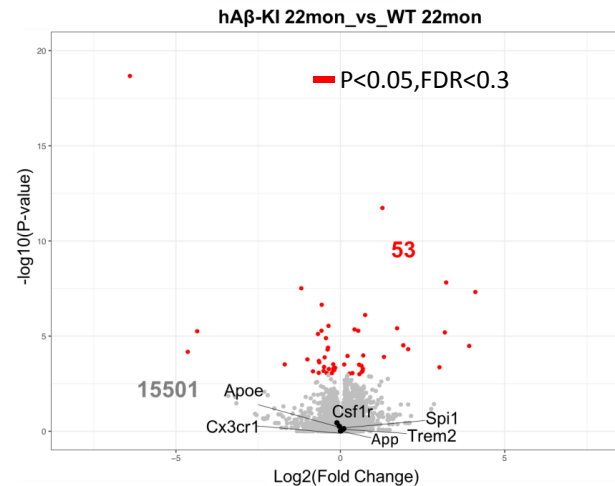
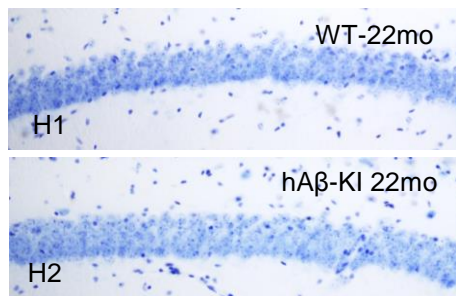
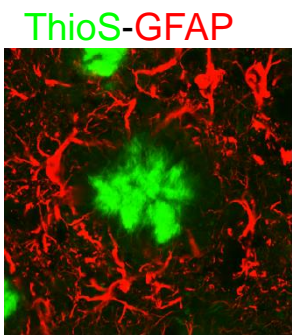
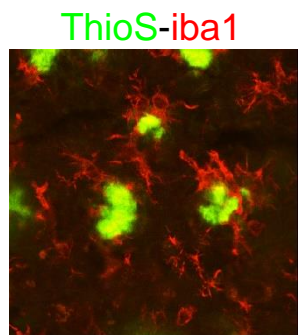
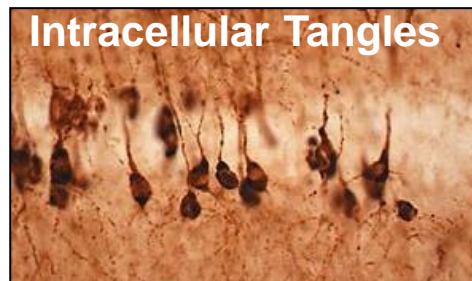


Deep phenotyping pipeline for LOAD models – UCI DMP

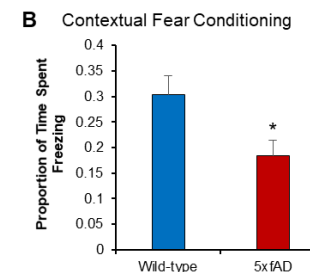
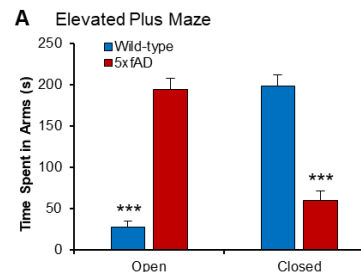


Model Characterization at UCI – 4,8,12,18 month timepoints

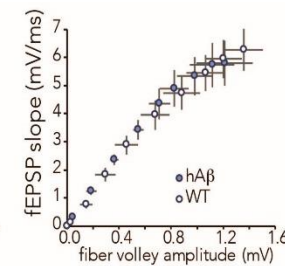
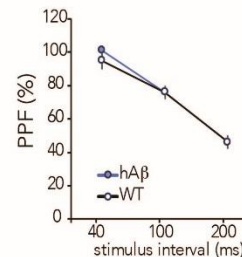
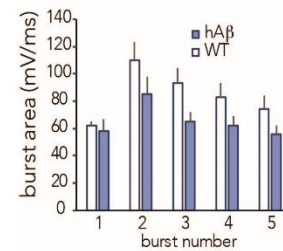
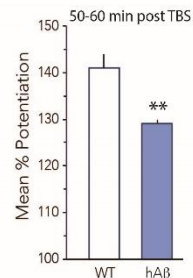
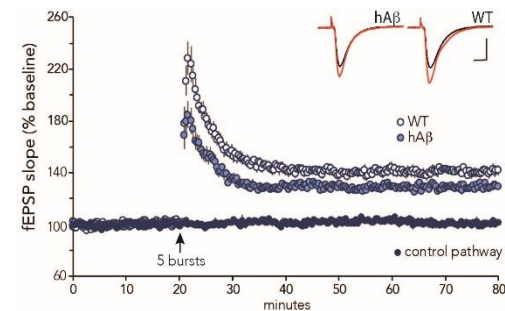
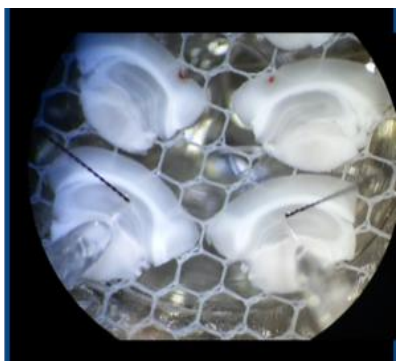
Neuropathology and Neurodegeneration



Network analysis:
Molecular Profiling
(RNA-Seq)



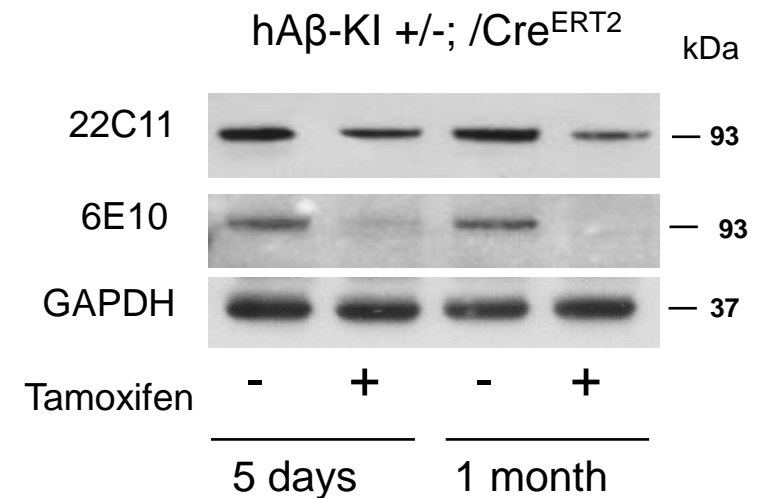
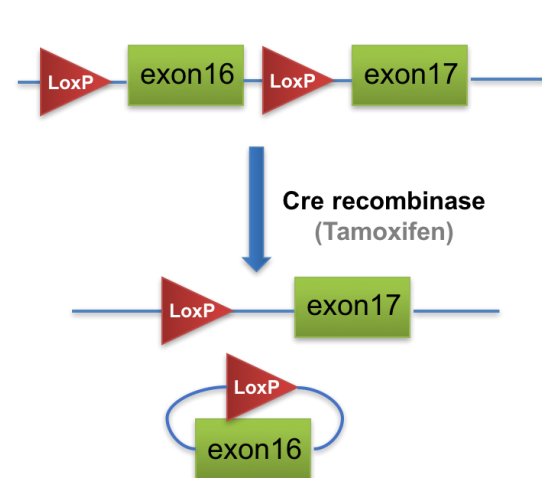
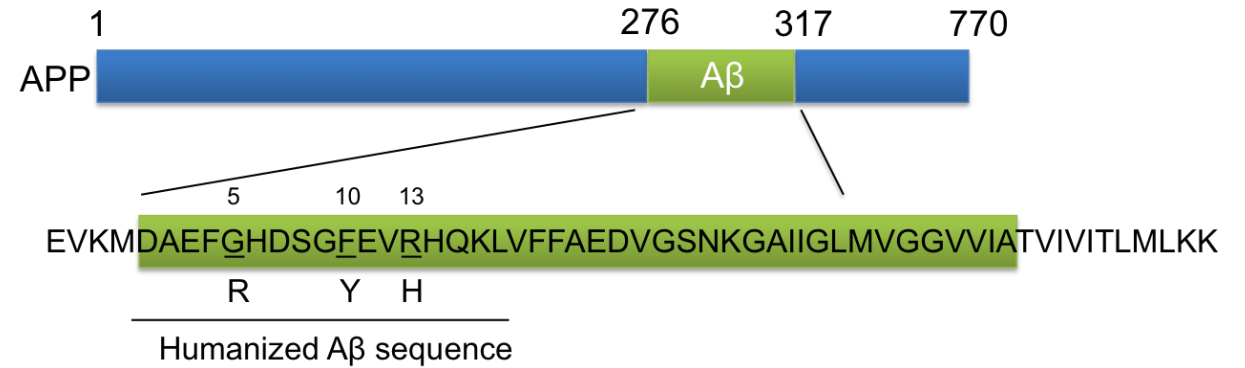
Behavioral and
Cognitive
Phenotyping



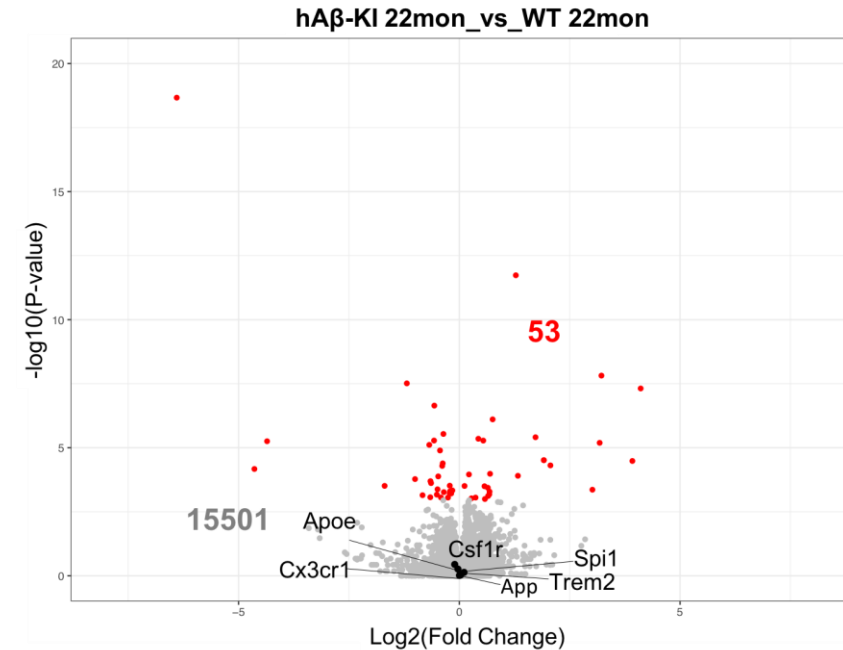
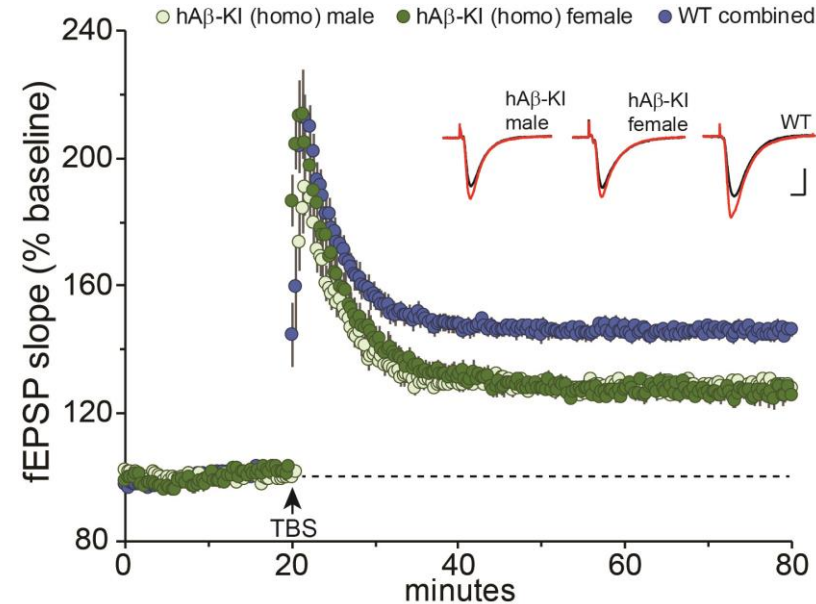
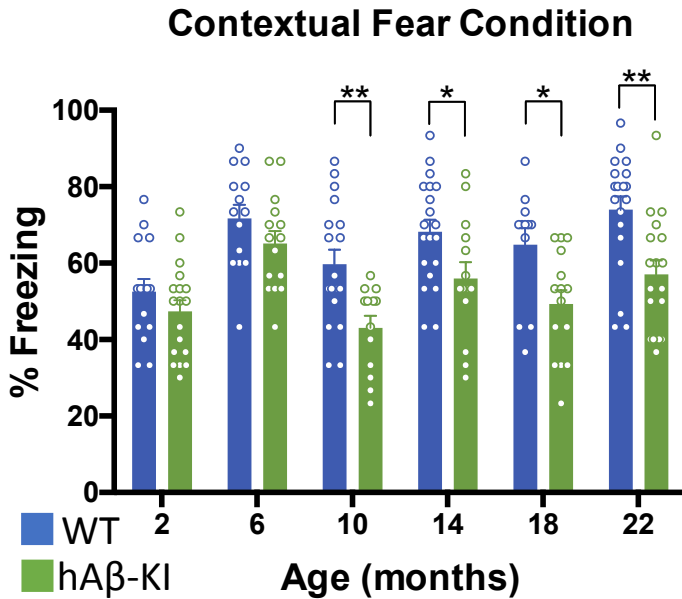
Electrophysiology

Generation of mice expressing a *cre-loxP* conditional allele of humanized wild-type A β □ hA β -loxP-KI model – UCI DMP

- No published allele of mouse *App* expresses normal human A β .
- Exon 16 humanized A β sequence is floxed, enabling cre-mediated cKO of humanized allele.
- IU/JAX has generated mice with complementary hA β -KI allele without *loxP* sites.
- Important models to investigate inherent difference in A β biology, plus provide platform for LOAD modeling.

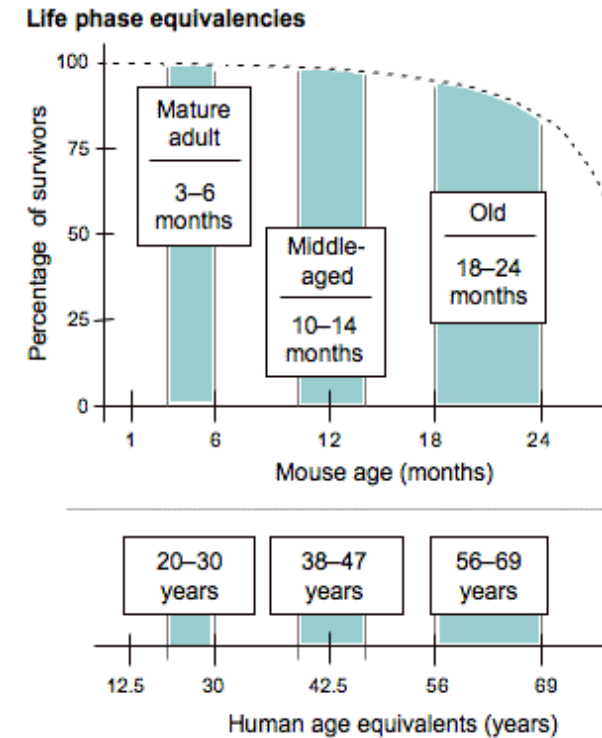
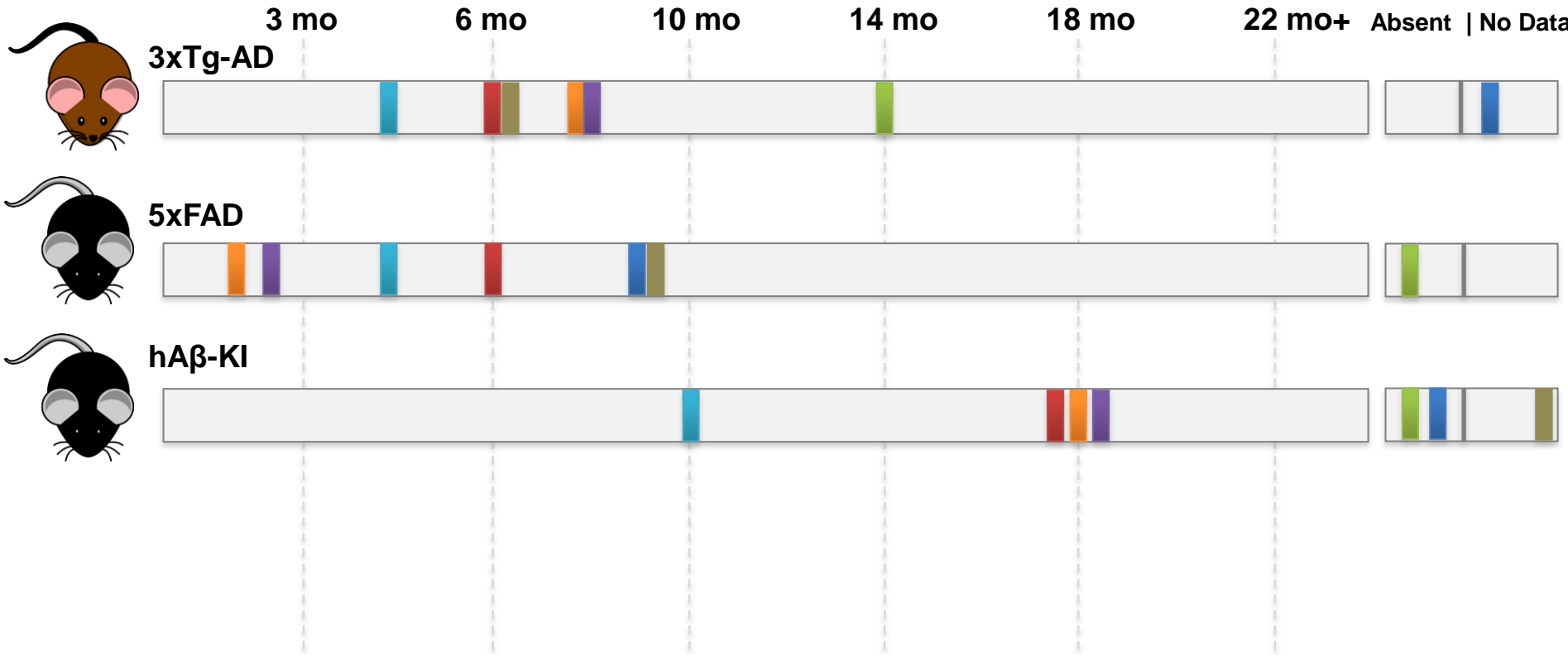
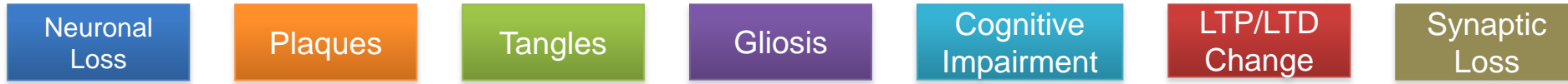


Mice expressing humanized wild-type A β display age-related altered cognition, electrophysiology and gene-expression



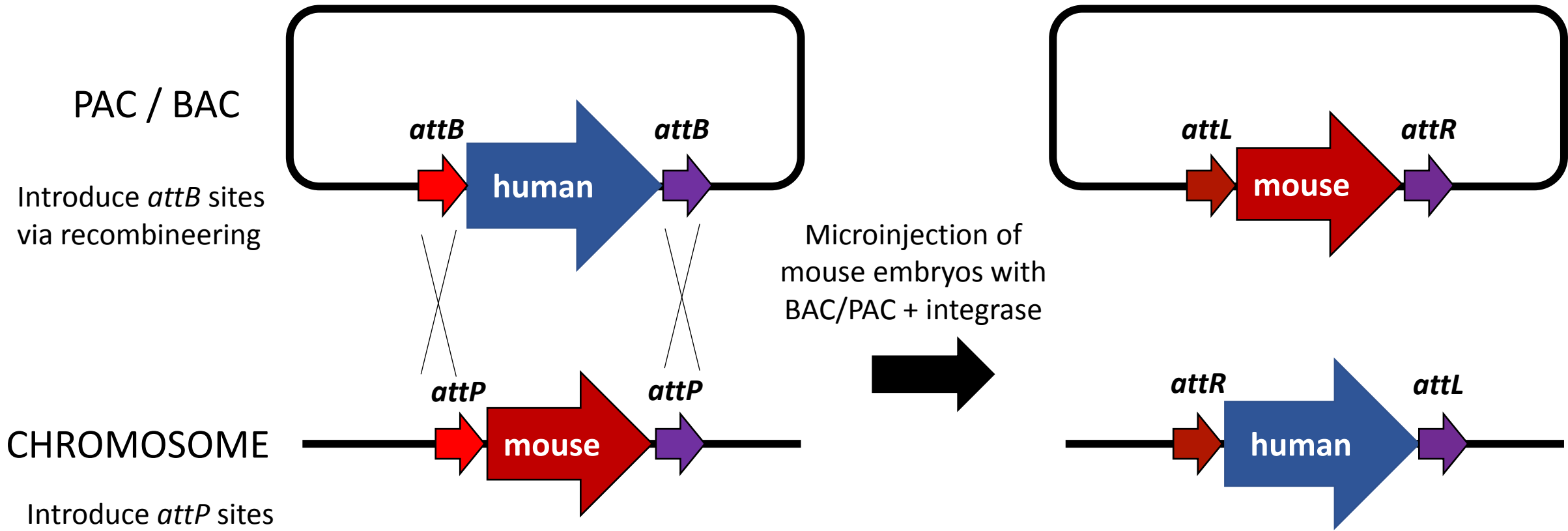
| Program, Approved Format, and original ID | Session Name, Date, Time | Presentation Title | Accept | Decline | Role | Presentation Time |
|--|--|---|----------------------------------|-----------------------|-------------------|-------------------|
| Alzheimer's Association International Conference (AAIC) | O1-01 Development of New Models and Analysis Methods: Novel Model Systems to Study Dementia, <i>Sunday, July 22, 2018: 8:00 AM - 9:30 AM, McCormick Place, Room - 184</i> | O1-01-04 Ha β -KI: A Knock-in Mouse Model for Sporadic Alzheimer's Disease | <input checked="" type="radio"/> | <input type="radio"/> | Presenting Author | 8:45 AM-9:00 AM |
| | | | David Baglietto-Vargas | | | |

Timeline for Development of Pathology in Mouse AD MODELS

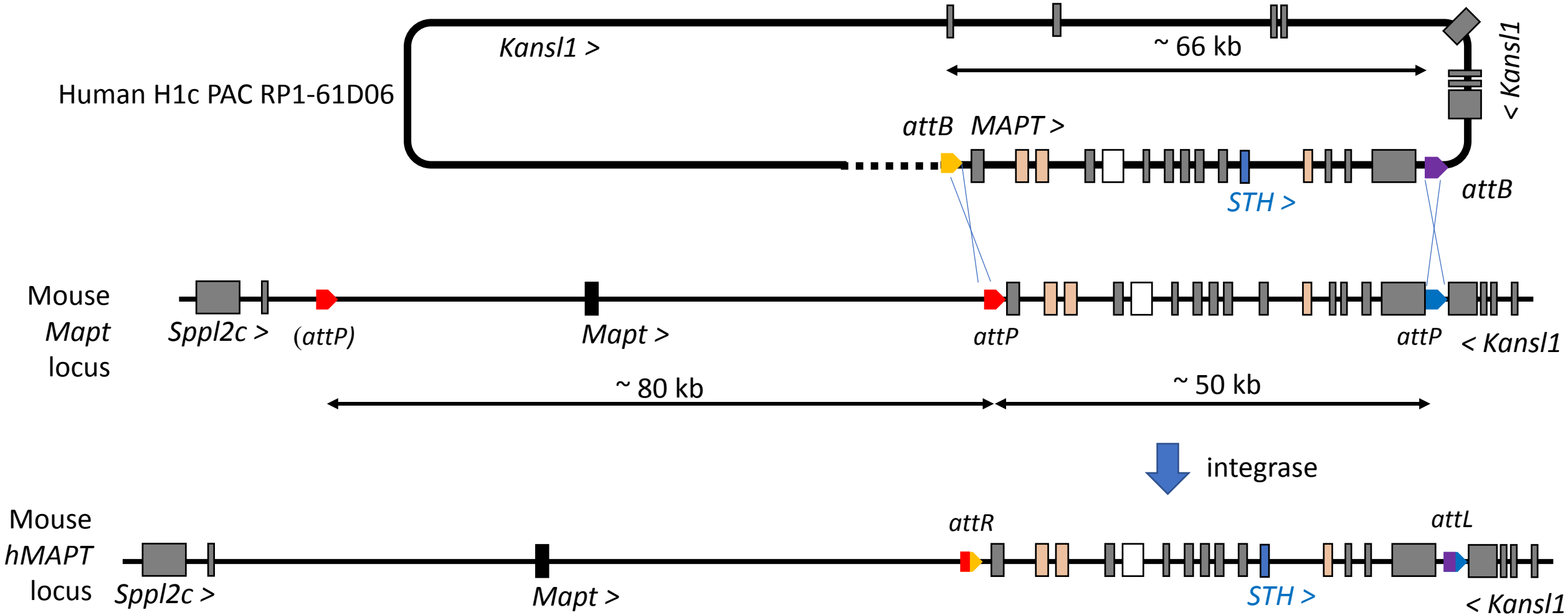


Flurkey K, Curren JM, Harrison DE. 2007.

Strategy to humanize mouse *Mapt* (TAU), *Clu* and other loci using Recombinase Mediated Cassette Exchange (RMCE)



hTau-KI mice - humanization of mouse *Mapt* via RMCE



IU/JAX Disease Modeling Project: 40 new models of LOAD



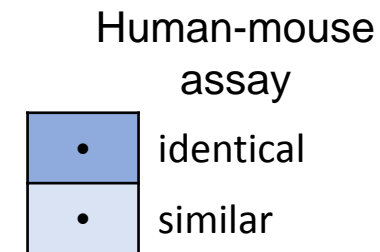
B6J.APOE^{E4/E4}TREM2^{R47H/R47H}
Common name: B6J.hAT

Early goals of IU/JAX DMP

- Characterize commonly used EOAD models
 - *APP/PS1* (Borchelt)
 - 5xFAD (Vassar)
 - hTau (Davies)
- Characterize newly created B6J.hAT LOAD model
- Introduce known GWAS human variants into *APOE/TREM* ‘sensitizer’ strain.
- Characterize and stage F344-Tg(PrP-APP, PrP-PS1) – rat model of EOAD

Clinically-relevant deep phenotyping

| AMP-AD, ADNI etc. | MODEL-AD | |
|---------------------------------|---|---|
| | Primary Screening 2, 6, 12 months 24 models | Deep Phenotyping 4, 8, 12, (18 months) Prioritized models |
| Assay | | |
| Amyloid and tau pathology | • | • |
| Neuroinflammation | • | • |
| Neuronal cell loss | • | • |
| Biomarkers | • | • |
| Biomarkers (Quanterix) | | • |
| Transcriptomes (NanoString) | • | |
| Transcriptomes (RNA-seq) | | • |
| Transcriptomes (scRNA-seq) | | pilot study* |
| Proteomics | | pilot study* |
| Metabolomics | | pilot study* |
| Imaging (FDG, PET/MRI) | | • |
| Cognitive tests | | • |



Pilot studies

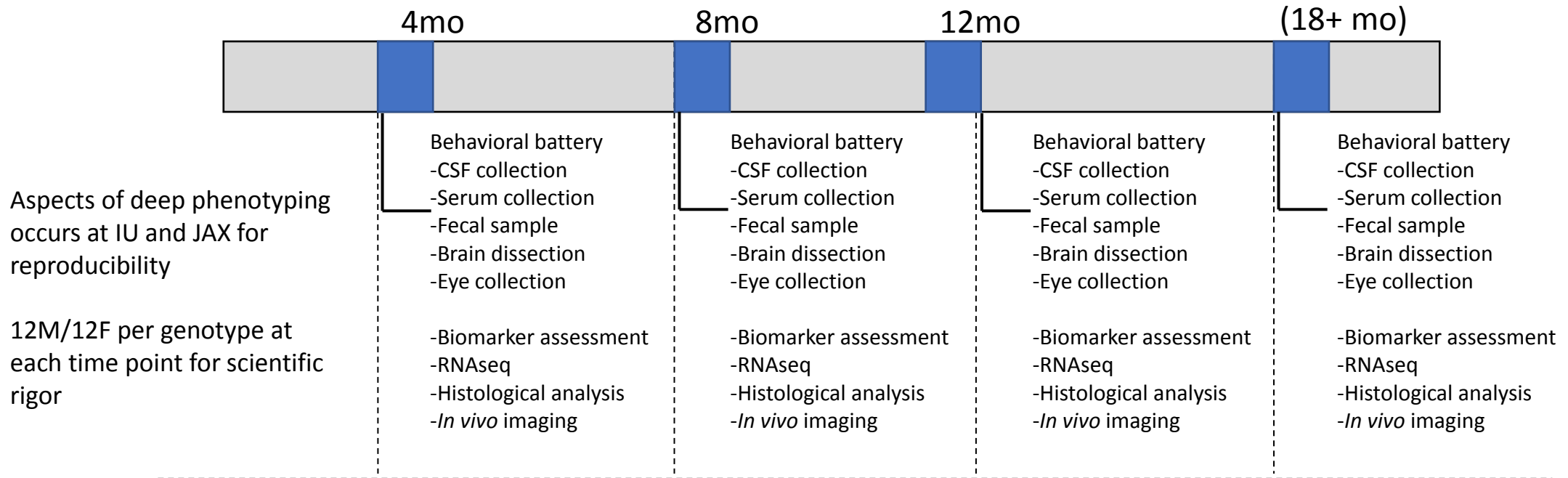
using B6J.5xFAD and B6J.*hAT*

scRNA-seq:
de Jager

Proteomics:
Seyfried

Metabolomics:
Kaddurah-Daouk

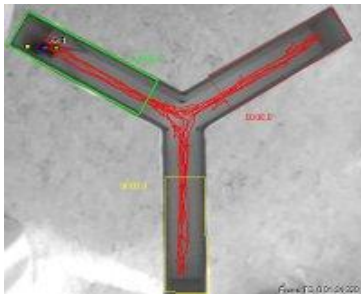
Clinically-relevant deep phenotyping



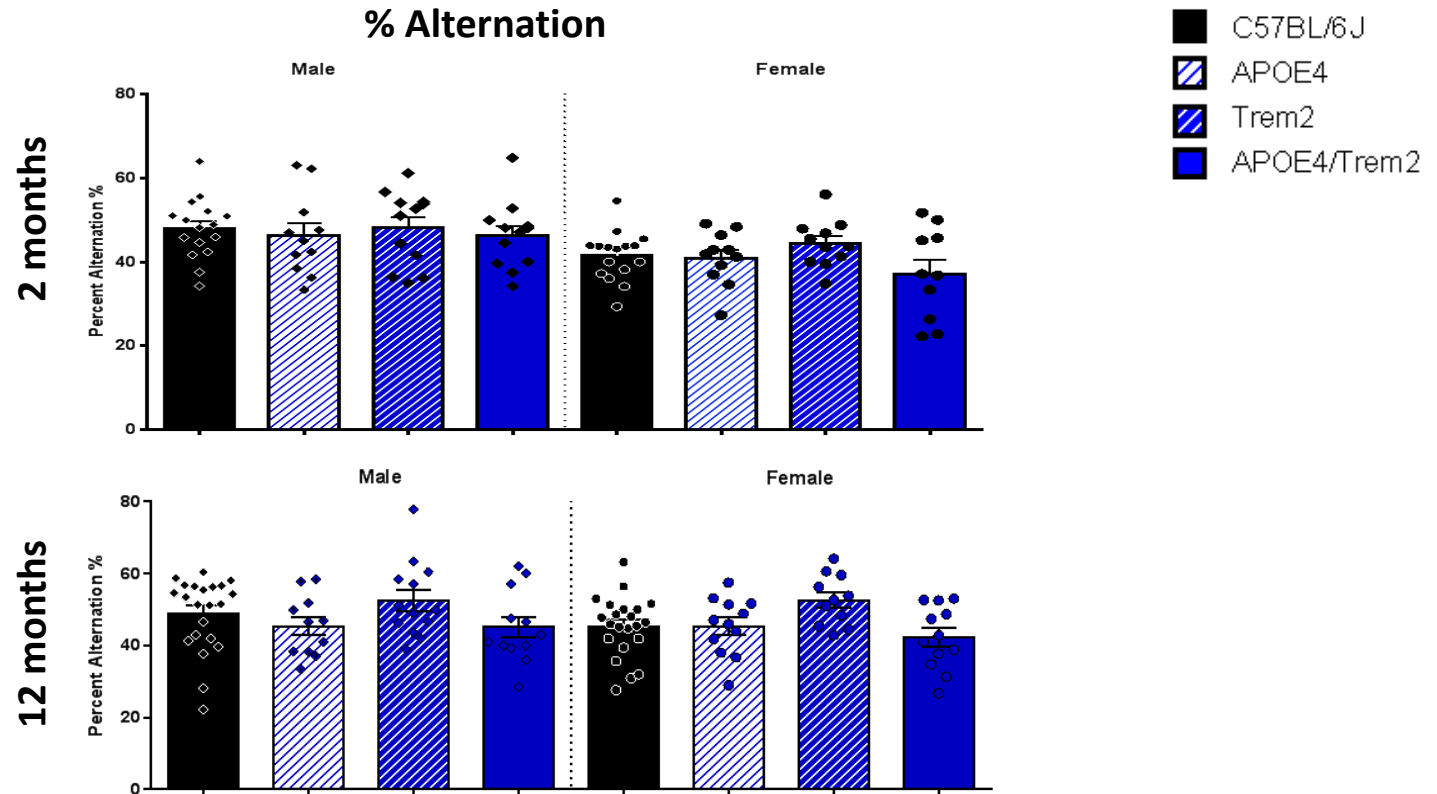
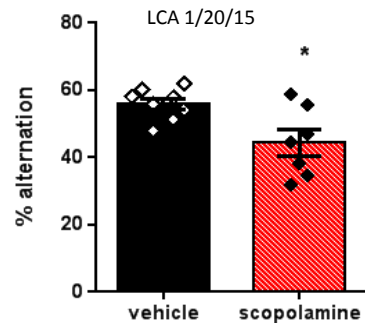
| | | | |
|---|--|---|---|
| <p>In vivo imaging by MR/PET: Amyloid: 18F-AV45 Tau: 18F-1451 Glucose: 18F-FDG Blood flow: 64Cu-PTSM</p> | <p>Biomarkers: AB, Tau Nfl Neurogranin sTREM2</p> | <p>Histology: Gross morphology/white matter: Luxol fast blue and Cresol Violet Neurons: NeuN and CTIP Plaques, dystrophic neurites and myeloid cells: X34, LAMP1 and IBA1 TAU: AT8 and H&E Neuroinflammation: IBA1 and GFAP Vascular health: CD31 and IBA1</p> | <p>Pilots: Proteomic and metabolomics profiling Considering: Microbiome</p> |
|---|--|---|---|

B6J.hAT: No differences in hippocampal working memory between genotypes and ages

Spontaneous Alternation



Task validation

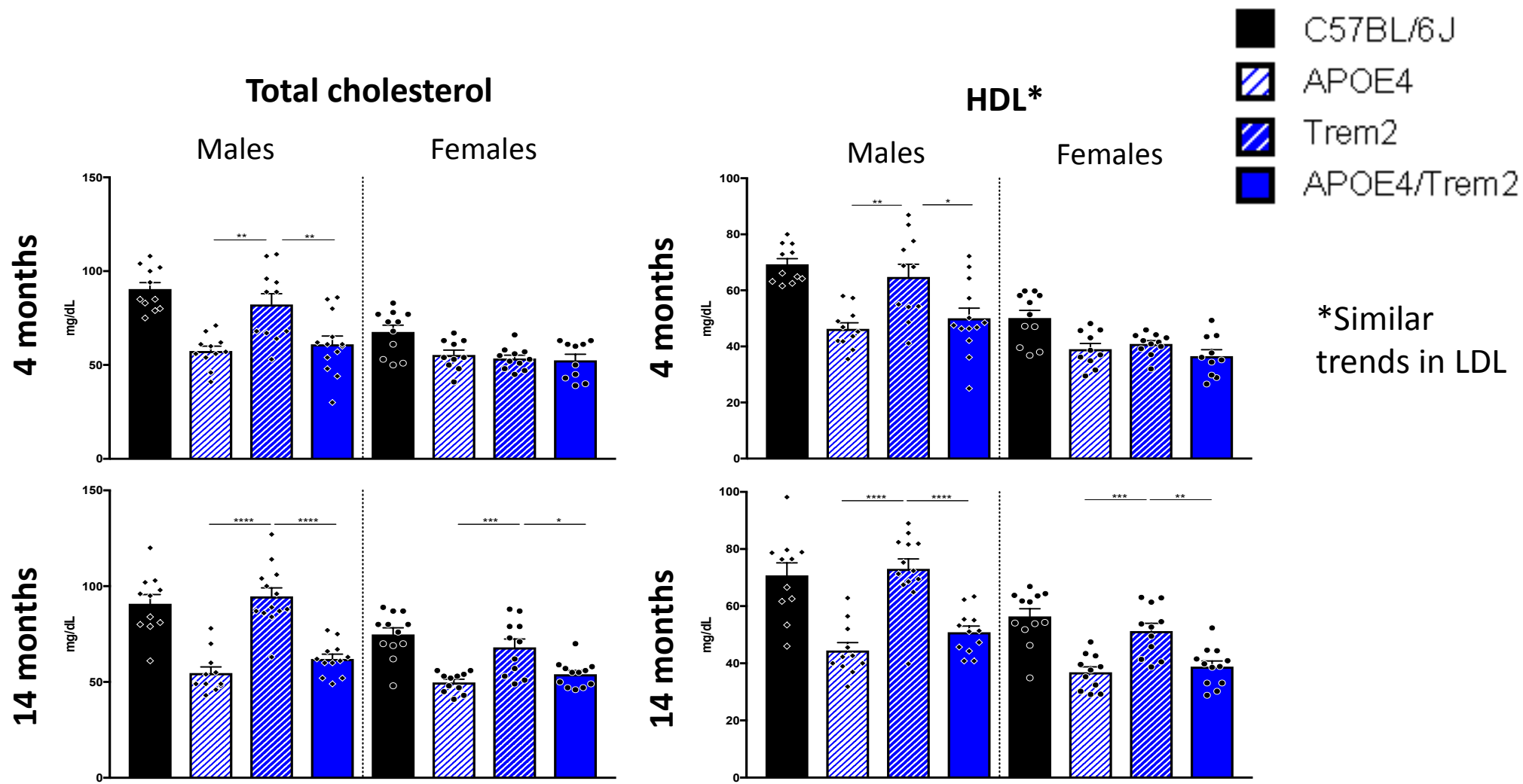


B6J.hAT: Differences in lipid profiles driven by *APOE*^{E4}

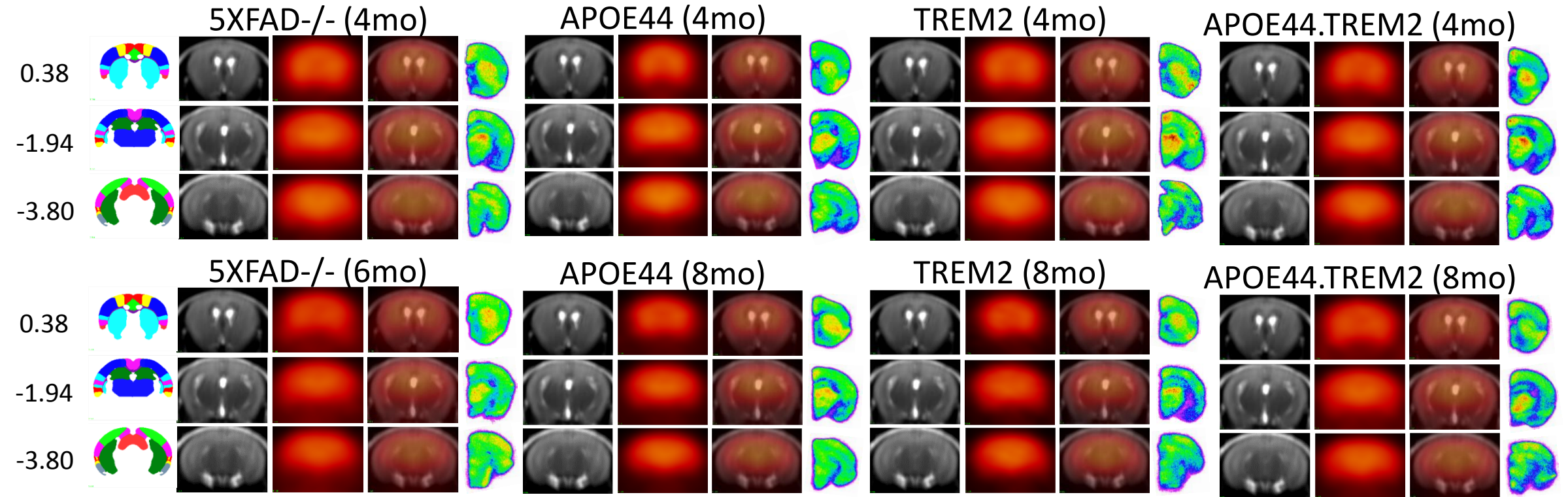
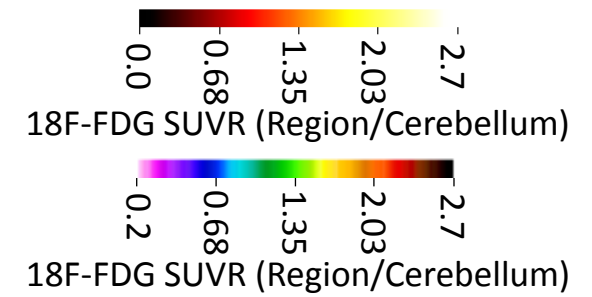
Blood collected at harvest (non-fasted)

Assessed for:

Total Cholesterol
LDL
HDL
Triglycerides
Non-essential FA
Glucose



B6J.hAT: PET/MR imaging (3T) with 18F-FDG



Preliminary findings: No significant changes across 27 brain regions comparing all genotypes at 4 and 8 mos

B6J.hAT Summary and Future plans

- *APOE*^{E4}-dependent changes in lipid profiles
- No age-dependent decline in glucose uptake across all genotypes
- No evidence of cognitive decline (measured using spontaneous alternation) up to 12 months of age

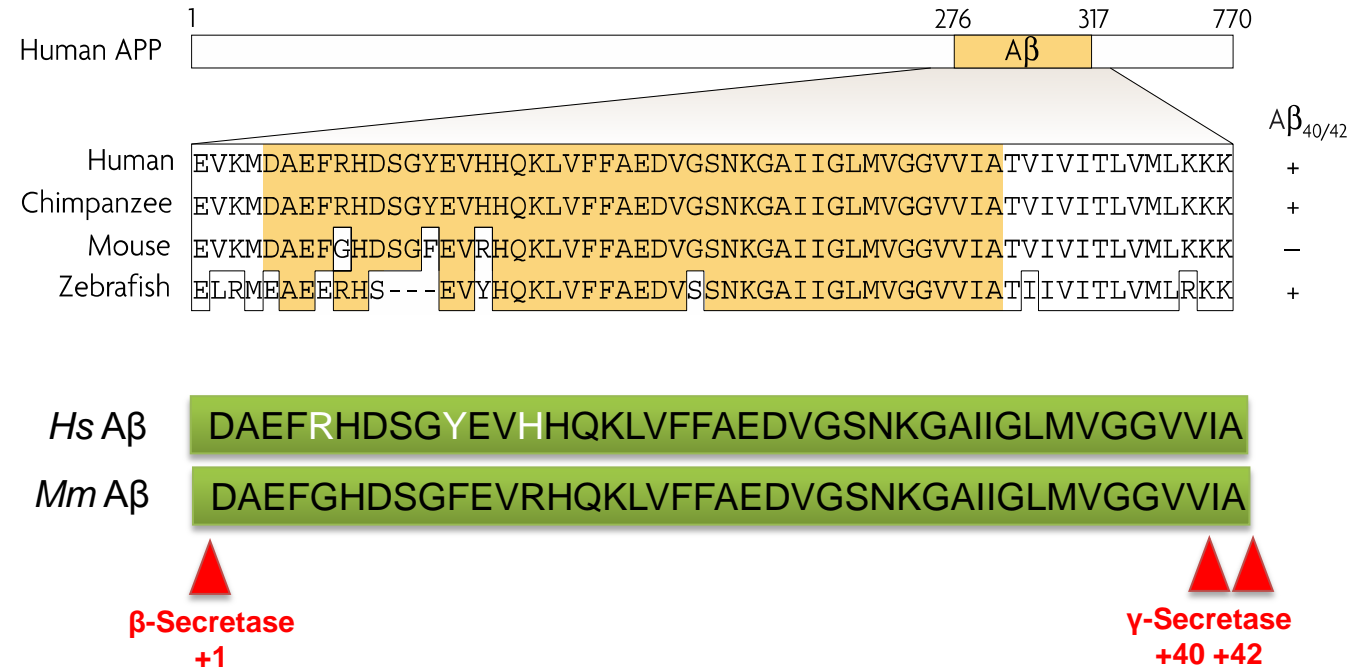
- Analyses of RNA sequence data of half brains underway
- Histological and biochemical assessment of tissue underway
 - Neuroinflammation, cerebrovascular health, amyloid and Tau

Next step: Deep phenotyping *B6J.APOE^{4/4}* *TREM2^{<R47H>}* mice with humanized *APP*



B6J.hATA

B6J.APOE^{E4/E4}TREM2^{R47H/R47H}App^{h/h}



Primary screen of novel variants on sensitized genetic background

Current



B6J.hAT

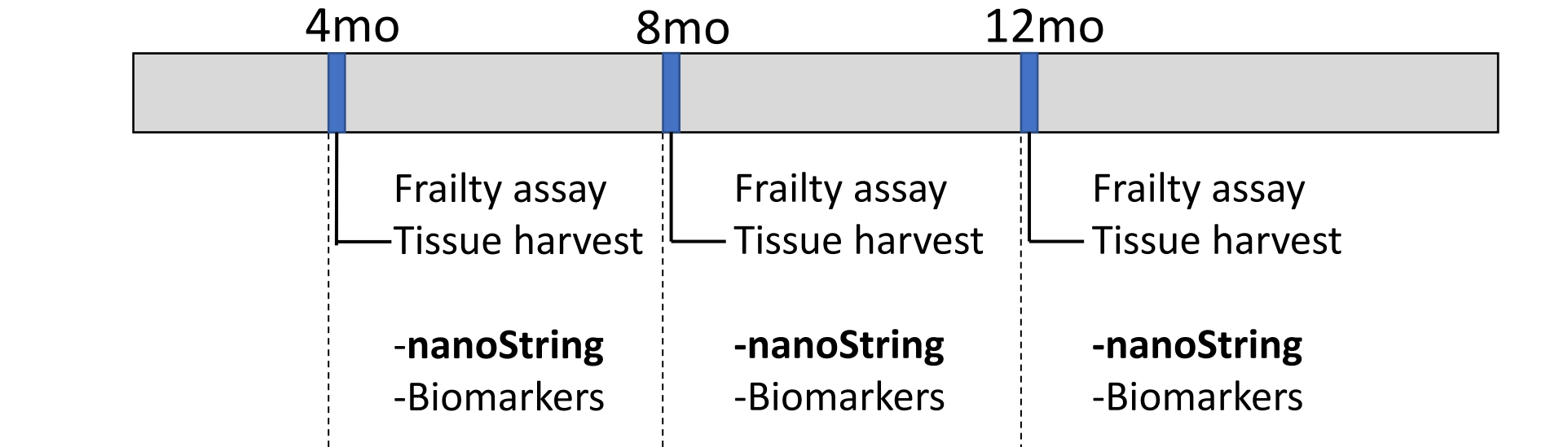
Future



B6J.hATA

- New strains created and in the primary screening pipeline
 - *Abca7*^{A1527G}, *Il1rap*^{KO}, *Ceacam1*^{KO},
Plcg2^{M28L}, *Mthfr*^{A222V}
- Ten others in CRISPR pipeline using *B6J.APOE*^{4/4} *TREM2*^{<R47H>}
- Up to 40 variants to be created with 24 to be screened

A primary screen to prioritize candidate variants for deep phenotyping



Promising strains prioritized for deep phenotyping

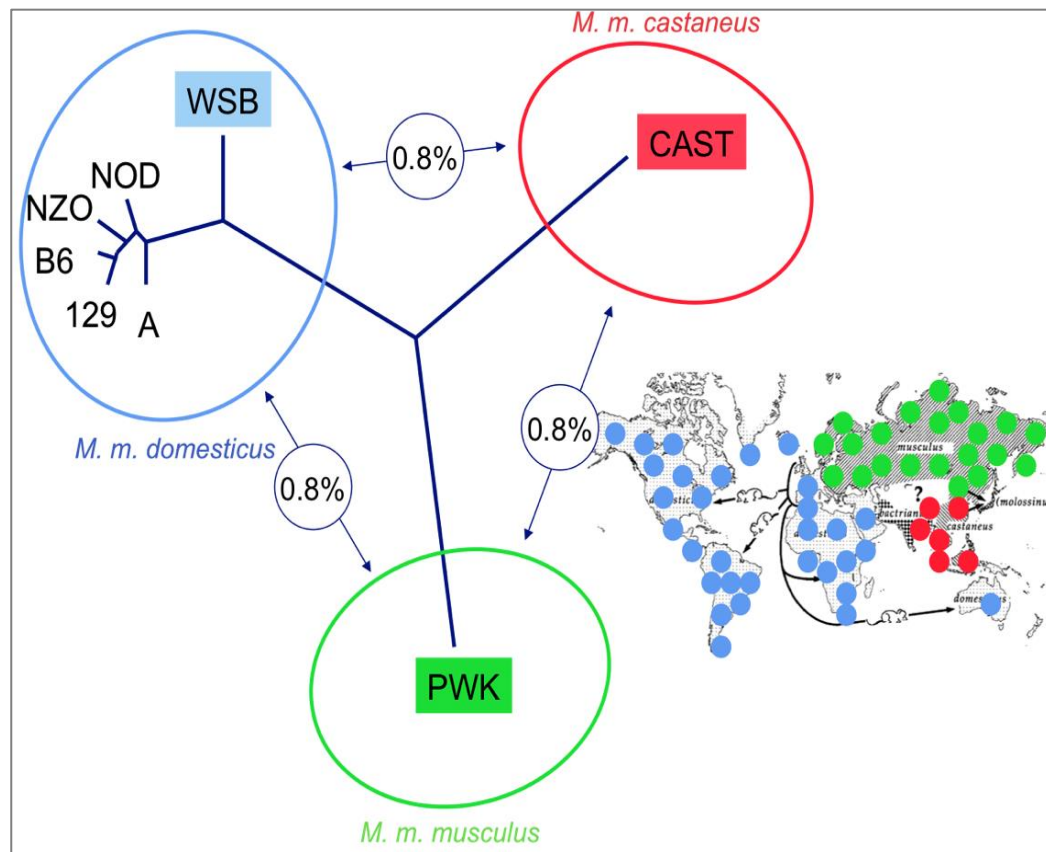
A nanoString panel to align mouse models to human data: AMP-AD panel

- Panel of 770+30 mouse gene probes
- Maximize coverage of 30 AMP-AD modules
- Include top AMP-AD candidates (Top 30, AGORA targets)
- Genes ranked by
 - representation of module PCs (gene score)
 - ortholog expressed in mouse brain at 6 months of age
 - 10 housekeeping genes

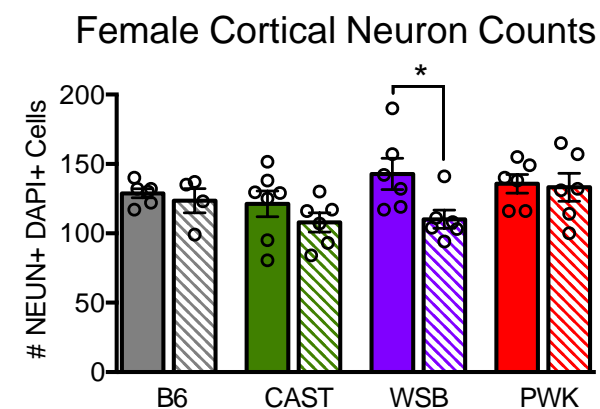
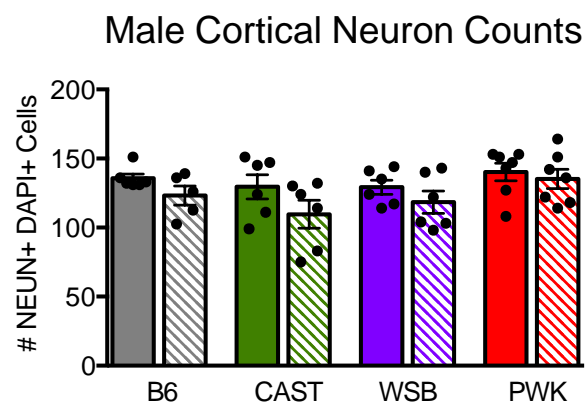
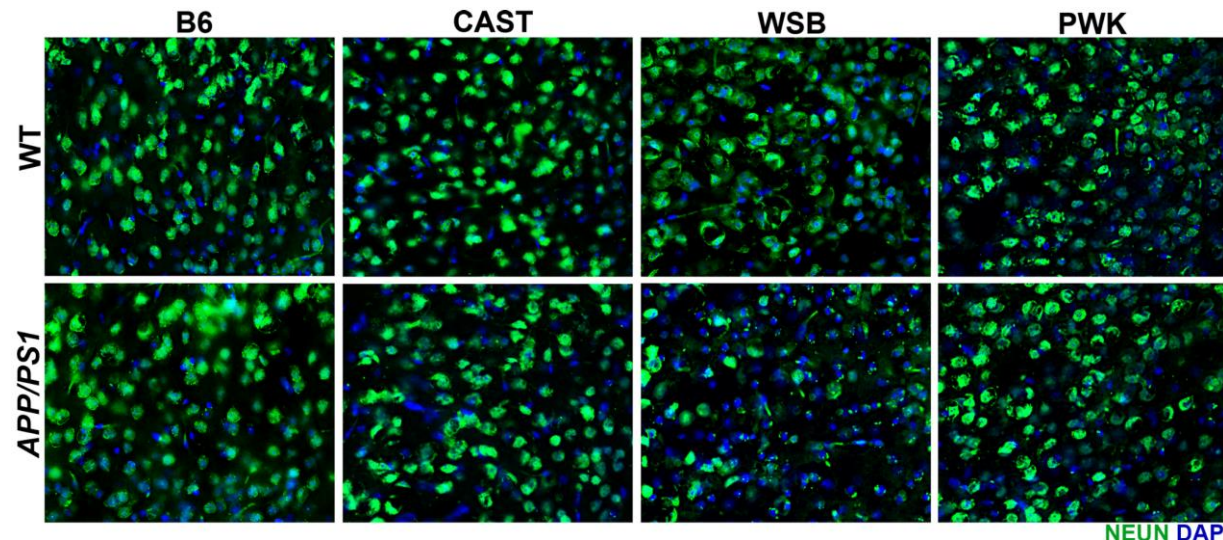
| AMP-AD Module | Nanostring probes per module | AMP-AD Module Size | % of Module Covered |
|-------------------------|------------------------------|--------------------|---------------------|
| aggregateCBEblue | 177 | 4505 | 3.93 |
| aggregateCBEbrown | 95 | 504 | 18.85 |
| aggregateCBeturquoise | 200 | 1977 | 10.12 |
| aggregateCBEyellow | 157 | 1738 | 9.03 |
| aggregateDLPFCblue | 183 | 1751 | 10.45 |
| aggregateDLPFCbrown | 139 | 882 | 15.76 |
| aggregateDLPFCturquoise | 144 | 2489 | 5.79 |
| aggregateDLPFCyellow | 192 | 3016 | 6.37 |
| aggregateFPblue | 278 | 1991 | 13.96 |
| aggregateFPbrown | 76 | 1287 | 5.91 |
| aggregateFPturquoise | 107 | 1001 | 10.69 |
| aggregateFPyellow | 188 | 4420 | 4.25 |
| etc | | | |

Coverage ranges from 76-278 genes per module

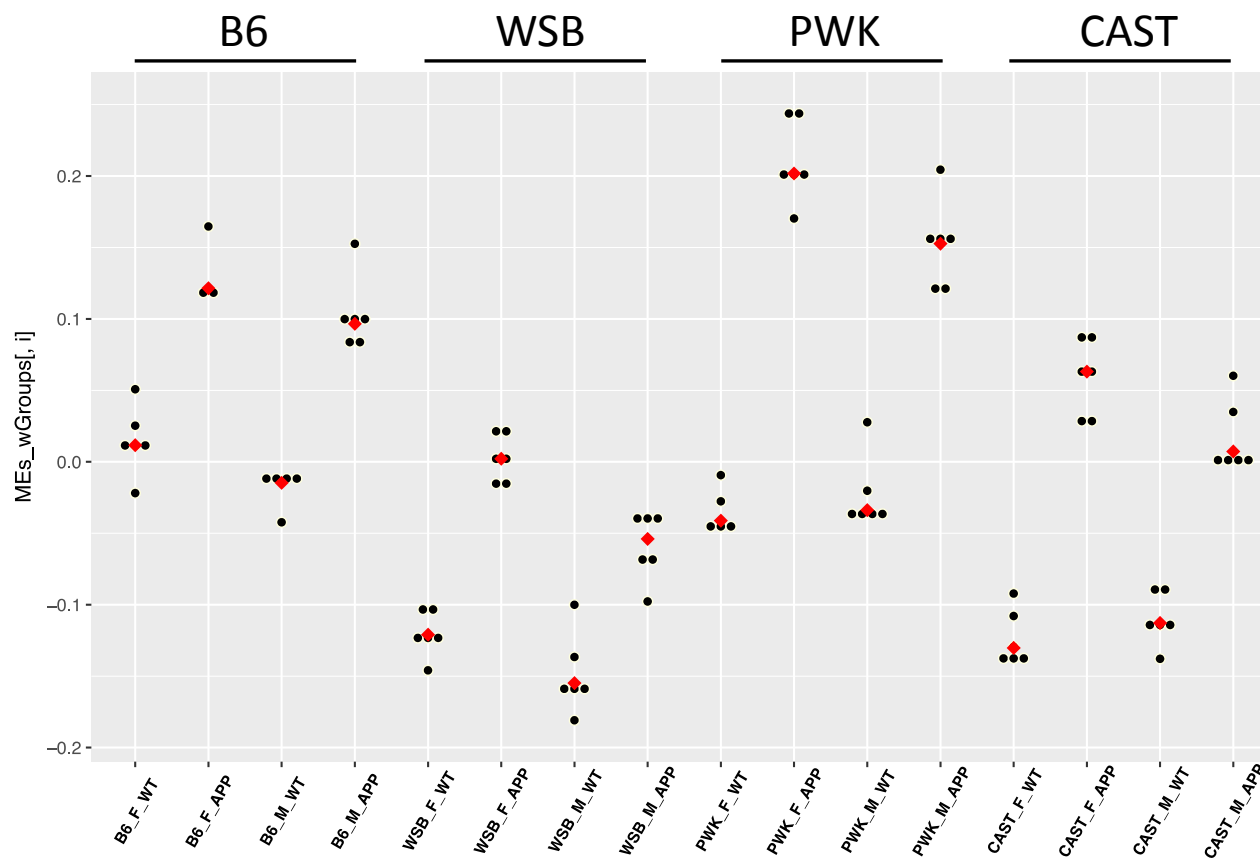
Genetic context is important



APP/PS1 on B6J and wild-derived strains



Transcriptome analyses by WGCNA shows variation in amyloid response between strains

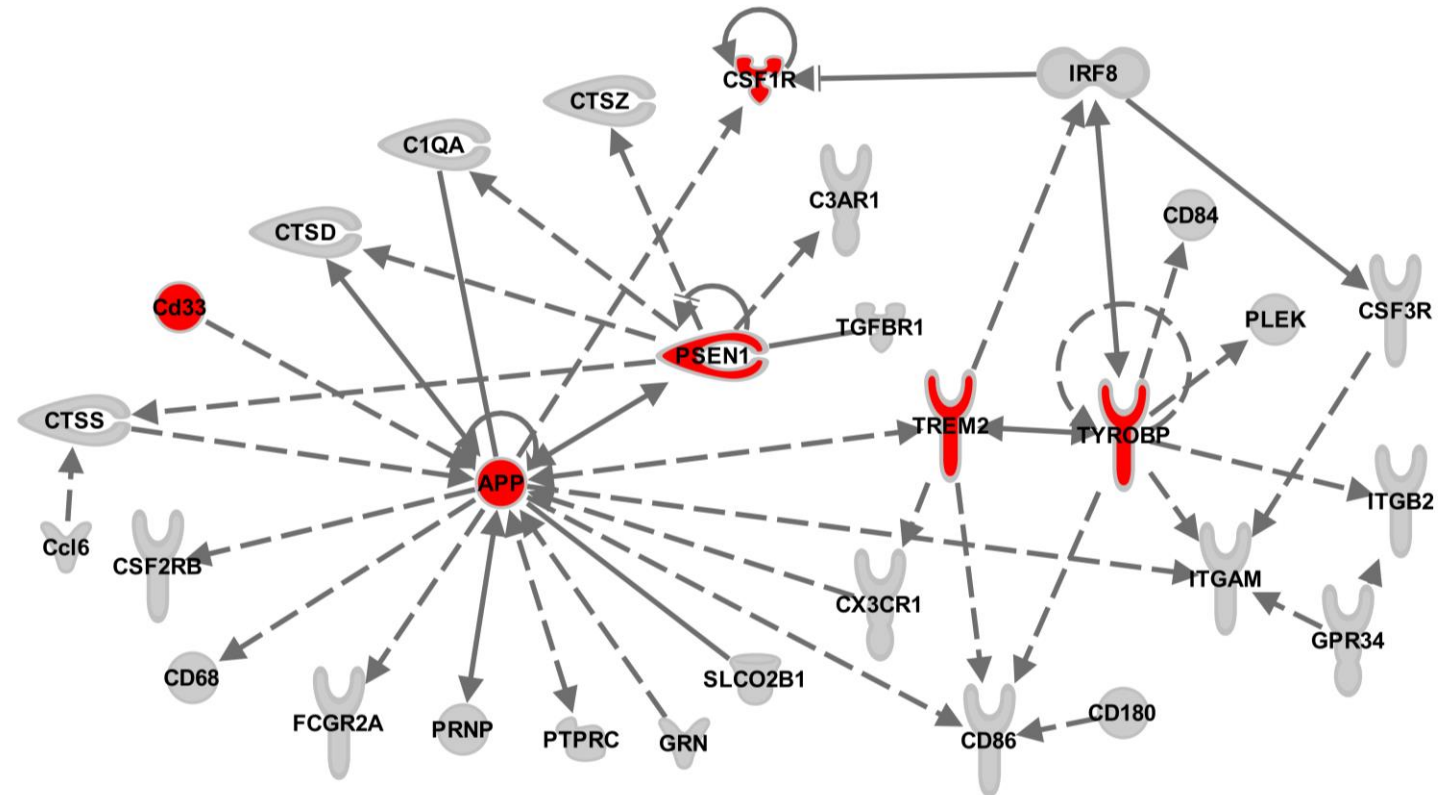
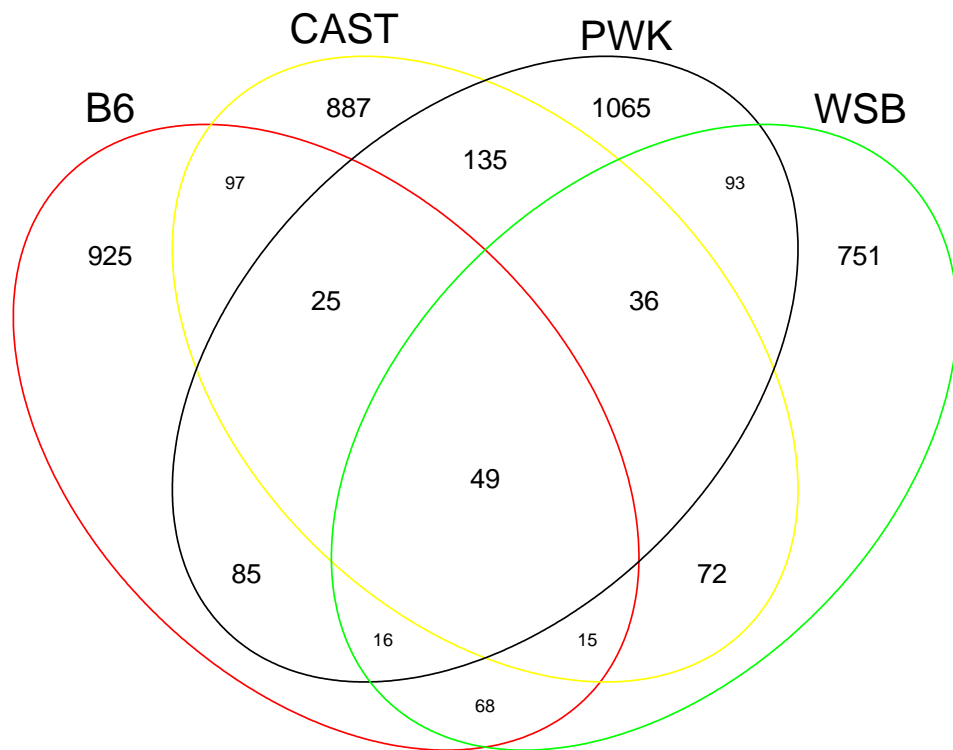


Genes in module

| | |
|--------|---------------|
| Itgb2 | Tbxas1 |
| Cd52 | Tyrobp |
| Spi1 | Tgfbr2 |
| Ptpn6 | Arpp21 |
| Ctsd | Vav1 |
| Ctsz | Cd84 |
| Abi3 | Ctss |
| Cd68 | Gpr34 |
| Cd180 | Cd53 |
| Fyb | Irf8 |
| App | Fam46c |
| Pros1 | Tlr7 |
| Trem2 | Mpeg1 |
| Csf1r | Gpr84 |
| Cndp2 | Csf2rb |
| Ptprc | Prnp |
| Slamf9 | |
| Lptm5 | 2900079G21Rik |

Strain-specific transcriptome analyses shows variation in genetic 'drivers' of AD

Generalized linear modeling



Summary: Evaluating the Translational Validity of Mouse Models of LOAD by clinically relevant deep phenotyping

- Creating up to 50 mouse models relevant to Alzheimer's disease
 - Includes creating a humanized platform (*APP*, *TAU*, *APOE*) for testing novel variants
 - Approximately 20 models created or in progress including AB-KI, hAT
- Perform clinically-relevant deep phenotyping of key (>10) models
 - Including in vivo imaging (MR/PET) and RNA-seq
 - Data available for 4 existing (5xFAD, 3xTG, APP/PS1, hTau) and 2 new models (hAB-KI, APOE4/TREM2<R47H>)
 - Pilots for proteomics and metabolomics underway
- All data and mouse strains made available through Synapse and JAX mouse repository (as well as other sources)
 - 23 models either available to order, available for preorder, or in preparation
 - ~265 RNA-seq data files submitted/being submitted to Synapse (many more to come!)

Strains and data available from model-ad.org

The screenshot displays the model-ad.org website interface. At the top, logos for Indiana University, The Jackson Laboratory, Sage Bionetworks, and UCI are visible. A navigation menu includes 'HOME', 'CORES', 'EXTERNAL ADVISORY BOARD', and 'RESOURCES'. The 'RESOURCES' link is highlighted with a red box and an arrow pointing to the 'Strain Table'.

Strain Table

| Category | Common name | Strain | Availability | Source |
|------------------------------|--------------------------------|----------------|--------------|----------|
| Late-onset AD-related models | APOE4/Trem2*R47H | B6j | live | MODEL-AD |
| familial AD models | APOE4 KI | B6j | live | MODEL-AD |
| 5XFAD | APOE3 KI | B6j | live | MODEL-AD |
| APP/PS1 | APOE2 KI | B6j | ~Fall '18 | MODEL-AD |
| 3XTg-AD | hAbeta-loxP-KI | mixed B6j; B6N | ~Fall '18 | MODEL-AD |

MOUSE STRAIN DATASHEET - 028709

B6(SJL)-Apo^{tm1.1(APOE*4)}Adiuj Trem2^{em1Adiuj}/J **NEW**

Stock No: 028709 | APOE4/Trem2*R47H

REPOSITORY LIVE

PLACE ORDER

3-6 week average lead time depending on quantity and age requests are not accepted

| | | | | |
|--------------------------|-----|---------------------------------------|------|----------|
| Trem2 KO | B6j | C57BL/6J-Trem2 ^{em2Adiuj} /J | live | MODEL-AD |
|--------------------------|-----|---------------------------------------|------|----------|

The MODEL-AD Consortium

Indiana University

Bruce Lamb, Program Director
Paul Territo, PTC Head
Andrew Saykin, BDMC Co-Head
Adrian Oblak, Project Manager
Kwangsik Nho
Li Shen
Tatiana Foroud
Dino Ghetti
David Jones
Sarah Quinney
Deborah DeBusk, Administrator

Sage Bionetworks

Lara Mangravite, BDMC Co-Head
Larsson Omberg
Ben Logsdon
Mette Peters
Solveig Sieberts
Yooree Chae

The Jackson Laboratory

Gareth Howell, DMP Head
Greg Carter, BDMC Head
Mike Sasner, DMP Co-Head
Stacey Rizzo, PTC Co-Head
Harriet Williams, Project Manager
Christoph Preuss
Asli Uyar
Yi Li
Ravi Pandey
Cai John
Nikhil Milind
Kristen Onos
Martha Abbott, Administrator

UC Irvine

Frank LaFerla, Program Director
Andrea Tenner, Program Director
Grant MacGregor, DMP Head
Ali Mortazavi, BDMC Head
Kim Green, DMP Co-Head
Marcelo Wood, DMP Co-Head
Stefania Forner, Project Manager
David Baglietto-Vargas
Shan Jiang
Shimako Kawauchi
Sherilyn Collins
Jonathan Neumann
Eniko Kramar
Edna Hingco
Dina Matheos
Maria Fonseca
Andrea Wasserman, Administrator

Contact

www.model-ad.org
modelad@iupui.edu
[@Model_ad_alz](https://twitter.com/Model_ad_alz)

National Institute on Aging

Suzana Petanceska
Lorenzo Refolo
U54 AG054345, U54 AG054349